



Molecular diagnostic and drug delivery agents based on aptamer-nanomaterial conjugates[☆]

Jung Heon Lee^{a,e}, Mehmet V. Yigit^{b,e}, Debapriya Mazumdar^{c,e}, Yi Lu^{a,b,c,d,e,*}

^a Department of Materials Science and Engineering, University of Illinois at Urbana–Champaign, Urbana, IL 61801, USA

^b Center for Biophysics and Computational Biology, University of Illinois at Urbana–Champaign, Urbana, IL 61801, USA

^c Department of Chemistry, University of Illinois at Urbana–Champaign, Urbana, IL 61801, USA

^d Department of Biochemistry, University of Illinois at Urbana–Champaign, Urbana, IL 61801, USA

^e Beckman Institute for Advanced Science and Technology, University of Illinois at Urbana–Champaign, Urbana, IL 61801, USA

ARTICLE INFO

Article history:

Received 8 October 2009

Accepted 3 February 2010

Available online 22 March 2010

Keywords:

DNA

Aptamer

Nanomaterials

Diagnostics

Biosensor

Drug delivery

Biomedical

Targeted therapeutics

ABSTRACT

Recent progress in an emerging area of designing aptamer and nanomaterial conjugates as molecular diagnostic and drug delivery agents in biomedical applications is summarized. Aptamers specific for a wide range of targets are first introduced and compared to antibodies. Methods of integrating these aptamers with a variety of nanomaterials, such as gold nanoparticles, quantum dots, carbon nanotubes, and superparamagnetic iron oxide nanoparticles, each with unique optical, magnetic, and electrochemical properties, are reviewed. Applications of these systems as fluorescent, colorimetric, magnetic resonance imaging, and electrochemical sensors in medical diagnostics are given, along with new applications as smart drug delivery agents.

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1. Introduction

Recent developments in nanostructured material synthesis and engineering have made a huge impact on a number of fields including nanoelectronics, photonics, biology, and medicine [1,2]. A main reason for such an impact is that every nanomaterial, such as metallic nanoparticles

[☆] This review is part of the *Advanced Drug Delivery Reviews* theme issue on "From Biology to Materials: Engineering DNA and RNA for Drug Delivery and Nanomedicine".

* Corresponding author. Department of Chemistry and Department of Materials Science and Engineering, University of Illinois at Urbana–Champaign, Urbana, IL 61801, USA.

E-mail address: yi-lu@illinois.edu (Y. Lu).

(NPs), quantum dots (QDs), carbon nanotubes (CNTs), and magnetic nanoparticles has unique physical and chemical properties that can be used for various applications. Of particular relevance to this review are optical, magnetic, electrochemical, thermal, catalytic, and mechanical signals generated by these nanomaterials, making them ideal candidates for signal transductions in medical diagnostics.

In order for nanomaterials to be used as clinical probes, it is necessary to provide them with target recognition capability; this task is generally accomplished by functionalizing the nanomaterials with biomolecules. Proteins such as antibodies are well known targeting molecules to be conjugated onto the nanomaterials. Recently functional nucleic acids are an emerging class of molecules to be used together with nanomaterials for biomedical applications [3–6].

Functional nucleic acids are DNA or RNA molecules that can interact with or bind to a specific analyte, resulting in a conformation change or a catalytic reaction [7,8]. Since DNA is more stable and cheaper to produce than RNA, this review is mainly focused on functional DNA conjugated nanomaterials. Functional DNA molecules include DNAzymes, aptamers, and aptazymes. DNAzymes (also called catalytic DNA or deoxyribozymes elsewhere) are DNA molecules that can catalyze many chemical and biological reactions in the presence of specific molecules, mostly metal ions as cofactors. Aptamers, on the other hand, are nucleic acid molecules that can specifically bind to chemical or biological molecules. Aptazymes are the combination of DNAzyme and aptamer molecules. Once functional DNAs are combined with nanostructured materials, they can endow the nanomaterials with their target recognition capability allowing the hybrid system to be used as a sensor or agent for imaging/targeted drug delivery.

The target recognition ability of functional DNA has already been used extensively with nanomaterials to develop biosensors [9–11] for various target molecules to detect DNA [12–14], RNA [15], and metal ions [3,16–30]. Among them, aptamers modified with nanomaterials have great potential to be used for clinical diagnostics as they can detect diverse targets ranging from small molecules and proteins to intact viruses and cells.

In this review, various nanomaterial-aptamer systems for potential application in clinical diagnostics will be covered. First, aptamers will be introduced and their advantages and disadvantages will be discussed in comparison with antibodies. Following that, methods of combining aptamers with different nanomaterials to produce diagnostic agents, such as fluorescence, colorimetric, surface enhanced Raman scattering (SERS), magnetic resonance imaging (MRI), and electrochemical detection, will be summarized. Finally, recent progress in using the aptamer-nanomaterial conjugates for targeted drug delivery will be discussed.

2. Overview of aptamers

2.1. Aptamer

Aptamers can be considered as nucleic acid analogue of antibodies; they can bind with high affinity and specificity to a broad range of targets, such as small molecules, proteins, viruses, or cells. The concept of nucleic acid binding to target molecules or proteins was first introduced when a RNA with high affinity and selectivity to viral or cellular proteins was observed during the investigation on HIV and adenovirus in 1980's [31]. Aptamers can possess strong affinities to target molecules, with dissociation constants down to nanomolar or picomolar range. Currently, nucleic acid aptamers have been selected for more than 150 targets [32], including small molecules such as cocaine [33] and aspartame [34], growth factors [35–38], peptides [39,40], toxins [41,42], viral proteins [43,44], and cells and bacteria [45–47]. Among them some aptamers received approval from the US Food and Drug Administration (FDA) for vascular endothelial growth factor (VEGF)-165 isoform [48] and ocular vascular disease [49]. Recently, targeted drug delivery using polymeric nanoparticle (NP) and aptamer

conjugates on prostate cancer cells has been demonstrated *in vivo* [50], showing that aptamers have great potential in therapeutics and diagnostics [31,48,49,51–57].

2.2. *In vitro* selection of aptamer

Aptamers can be obtained via isolation from synthetic combinatorial libraries of nucleic acids through an *in vitro* selection process called SELEX (systematic evolution of ligands by exponential enrichment) [51,58,59]. The selection process starts with generating random sequences of 10^{14} – 10^{15} oligonucleotide strands which are chemically synthesized and amplified with polymerase chain reaction (PCR) (Fig. 1). The nucleic acid library is incubated with a target molecule, often immobilized onto a solid state matrix such as a gel or a column, so that the DNA or RNA having affinity to the target molecule can be captured. The target-bound nucleic acid molecules are separated from the unbound strands in the pool, following which the bound DNA or RNA strands are eluted from the target molecule and amplified via PCR to seed a new pool of nucleic acids enriched with sequences that have a higher affinity to the target. The next round of selection process is usually performed under more stringent conditions (such as lower target concentration and shorter time for binding). After ~ 10–20 rounds of the selection processes, the oligonucleotide with the highest affinity to the target molecule can be obtained. Furthermore, cell-based SELEX has been recently reported which enables to isolate aptamers directly from living cells [32,60]. This method allows selection of aptamers specific for the molecules present on the targeted cell. Since this work can be performed without the prior knowledge of the molecules on the cells, it not only allows early diagnostics but also opens opportunities for identification of new cancer cell markers.

2.3. Advantages of aptamers

Antibodies are being used in the majority of biomedical applications. Aptamers are an emerging class of molecules with several important advantages [51–55]. First, aptamers are efficient at binding not only to large molecules, such as proteins [61] and cells [62], but also to small molecules, such as nucleotides [63,64], organic dyes [65], amino acids [66], and metal ions [67,68], while antibodies are generally competent in binding to mostly larger molecules. Therefore there is a potential niche market for aptamers in diagnostics and drug delivery when small molecules are the targets. Second, aptamers are selected *in vitro*, and therefore can be used to select for a wide range of targets, including toxic or non-immunogenic molecules. Furthermore, the selection process can be performed under non-physiological condition including extremely high or low temperatures or pHs. These properties are difficult to be obtained with antibodies since they are produced *in vivo*. Third, once an aptamer is selected, it can be obtained in a large amount through chemical synthesis which is cost-effective and has minimal batch to batch difference in activity. It is easy to chemically modify aptamers with a variety of fluorophores, electrochemical or Raman reporters, or functional group such as primary amine, thiol groups on either 5' or 3' end, on the bases, or on the backbone of DNA [69]. Moreover, aptamers can be incorporated into one, two or three dimensional DNA-based nanostructures [70–75] which also have strong potential to be used for clinical and diagnostic applications [76–78]. In designing microarrays, aptamers can provide additional advantages. For example, as the size of aptamers is much smaller (~ 1–2 nm, < 10 kDa) than those of antibodies (~ 10 nm and ~ 155 kDa), aptamers have higher surface density [57] and less steric hindrance [51], which helps to increase the binding yield. In addition, non-specific adsorption happens much less for nucleic acid-immobilized surfaces compared to protein immobilized surfaces [79], which facilitates the engineering process. Finally, aptamers are much more stable to heat [80], pH [50], and organic solvents [81] than antibodies and, unlike antibodies, can be denatured and renatured

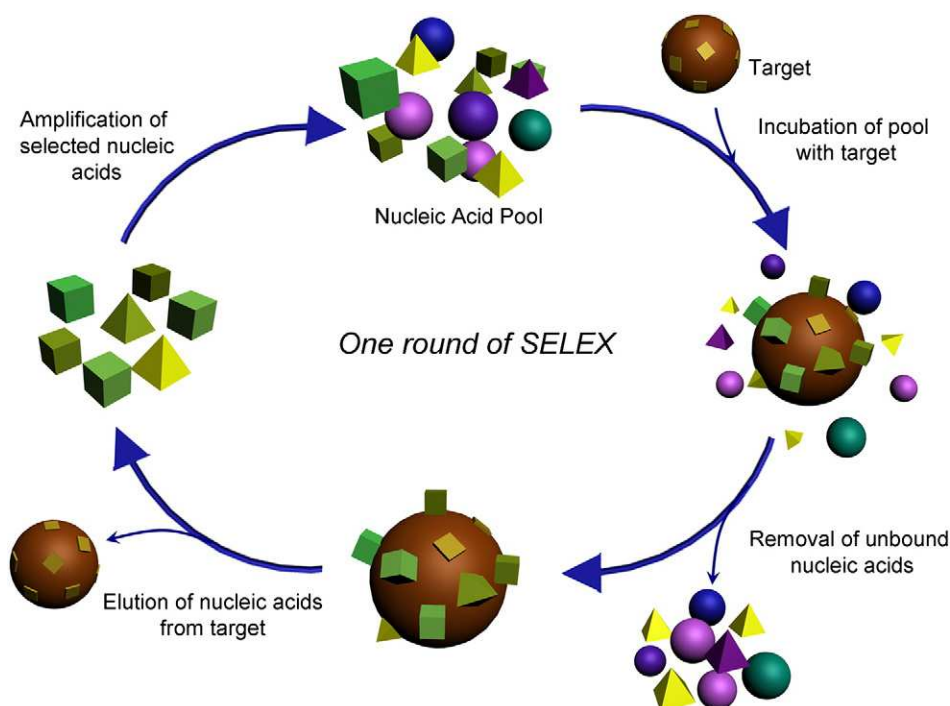


Fig. 1. Schematic depiction of SELEX. Nucleic acid pool containing 10^{14} – 10^{15} nucleic acids with random sequence is incubated with a target molecule. After removing unbound nucleic acids, nucleic acids are eluted from the target molecule. The pool for the next round is prepared after amplification and mutation of the selected nucleic acids. Selection can be completed after 10–20 rounds of selection process.

multiple times without significant loss of activity [82]. These properties make aptamers an excellent choice for biomedical applications.

2.4. Challenges facing aptamer-based diagnostics

Although aptamers are excellent candidates as diagnostic and drug delivery agents, there are still a few issues that have yet to be addressed for their practical applications. One issue is that nucleic acid aptamers could be vulnerable to nuclease degradation in cells or in blood [56]. This problem can be more significant for RNA aptamers because RNA molecules are much less stable to hydrolysis in biological fluid. To overcome this problem, several strategies have been proposed to increase the stability of aptamers, such as modification of aptamers with 2'-aminopyrimidine [83], 2'-fluoropyrimidine [83], or 2'-O-methyl nucleotides [84]. Another limitation aptamers might have is the necessity to limit its administration to local regions, since there is a possibility that proteins in untargeted organs can also be affected. However, some preliminary results showed the possibility of systematic delivery [51,52]. Finally, one of the major challenges is the development of general methods to convert the highly specific molecular recognition between aptamers and their targets into detectable signals. Conjugation of aptamers with nanomaterials can be an ideal way to overcome this hurdle.

3. Conjugation of nanomaterials and aptamers for biosensing and diagnostics

3.1. Fluorescence based sensors

Aptamers can be easily modified with a variety of organic dyes and thus fluorophore-based detection method has been the most widely demonstrated [3,5,6,85]. Since this review mainly focuses on the nanomaterial-based detection, fluorophore-based detection will not be discussed here.

Quantum dots (QDs) or semiconductor NPs are one type of fluorescent nanomaterials with several unique optical properties [86–88]. As compared to organic fluorescent dyes, quantum dots are

more photostable and the wavelength of the emitted light can be controlled by changing their size and composition of the materials. Furthermore, QDs have very broad excitation range but sharp emission, making it possible to excite different QDs with a single wavelength and yet result in variety of emission wavelengths. Based on these attractive properties, QDs have been broadly used for bioimaging and diagnostics.

QDs functionalized with DNA [89–91] have been used for the detection of DNA [92] and real time monitoring of hybridization procedure [93]. The first QD conjugated with aptamer was reported by Ellington and co-workers for energy transfer based detection of thrombin (see Fig. 2A) [94]. In this work, thrombin aptamers functionalized on QDs were hybridized with a complementary DNA strand labeled with a quencher at the end. In the absence of thrombin, the fluorescent signal of QDs was quenched because the QDs were placed in close proximity to the quenchers, resulting in the energy transfer from the quantum dot to the quencher. The presence of thrombin, however, recovered the signal as the thrombin-aptamer interaction induced the release of the complementary strand containing quencher.

A different type of QD-aptamer sensor was developed by Strano and co-workers (see Fig. 2B) [95]. In this work, they conjugated QDs with thrombin-specific aptamers. When thrombin was added, the interaction of thrombin with aptamers brought them close to QDs. Selective quenching of fluorescence occurred due to the charge transfer occurring from thrombin to the QDs. The sensor was very sensitive, with a detection limit of 1 nM, and had high selectivity.

For medical applications, it is advantageous to perform multiplexed detection of more than one analyte using a single sensor in an “one-pot” process. Our group reported multiplexed detection of adenosine and cocaine using both QDs and gold nanoparticles (AuNPs) conjugated with aptamers (Fig. 3) [96]. In this work, aggregates of QDs with emission peak at 525 nm and AuNPs were formed by DNA aptamer specific for adenosine. In parallel, aggregates of QDs with emission peak at 585 nm and AuNPs were formed by a cocaine specific DNA aptamer. While fluorescence from the sensor composed of the mixture of both assemblies was initially quenched due to energy transfer from each QD

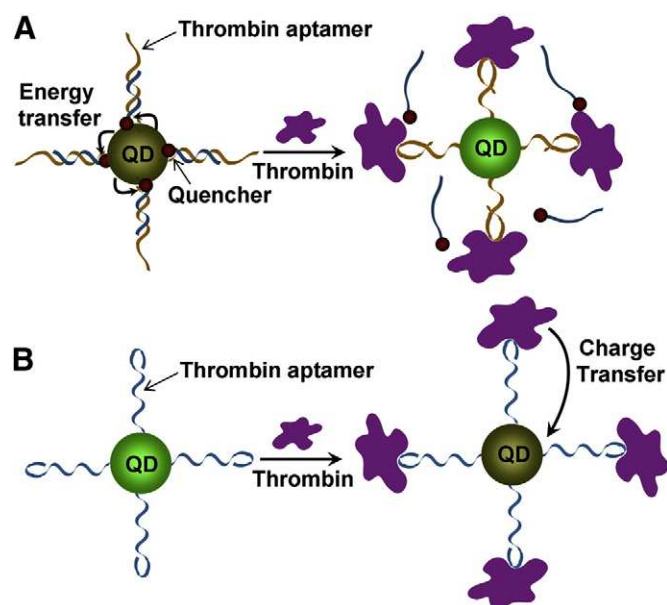


Fig. 2. Aptamer-QD based fluorescent sensors. (A) Thrombin aptamers are conjugated on QDs and hybridized to the complementary DNA with a quencher. Fluorescence of QD is quenched due to energy transfer from the QD to the quencher. The complementary DNA containing the quencher can be released after introduction of thrombin, inducing recovery of the fluorescence from QD. (B) Thrombin aptamers are conjugated to QDs. The fluorescence of QDs can be quenched as thrombin binds to aptamer due to the charge transfer from the thrombin to QD.

to AuNPs, the signal was enhanced at 525 nm with the addition of adenosine, or at 585 nm in the presence of cocaine as analyte-aptamer interaction induced disassembly of QD-AuNP aggregates. Fluorescence increase at both 525 nm and 585 nm was observed when both analytes were present.

It was recently reported that carbon nanotubes (CNT) also have fluorescence in the near-IR range which is advantageous for cell imaging by avoiding the high background fluorescence from the organelles in the cells [97]. Therefore, aptamer conjugated CNTs also have potential to be used as a fluorescence tag for cancer cell imaging. First, a photodynamic therapy agent generating singlet oxygen (1O_2) in the presence of α -thrombin was demonstrated using a photosensitizer labeled α -thrombin aptamer and single wall nanotube (SWNT) complex [98]. As the aptamer wrapped around the SWNT in the absence of target molecule, the labeled photosensitizer was placed close to SWNT, quenching the generation of singlet oxygen. On the other hand, the interaction between thrombin and aptamer induced release of photosensitizer from SWNT, generating singlet oxygen in the presence of light source. The same group further extended the methodology for detection of biomolecular interactions based on fluorescence [99].

Furthermore, fluorophore labeled DNA can also be immobilized on a solid surface. This provides several advantages such as low fluorescence background, regeneration, and long term storage compared to solution based fluorescence sensors [25]. In addition, the performance of fluorescence sensors can be engineered by the property of the material on which aptamer is immobilized. As the sensitivity and volume of the sample can be critical for some biomedical diagnostic applications, we investigated the influence of materials using Pb^{2+} specific DNAzyme labeled with fluorescein and Au surface as a model system. By immobilizing DNAzyme on a planar Au substrate, we could reach an order of magnitude improvement (1 nM) in the detection limit due to lower background, compared to its corresponding solution based sensor (10 nM) [100]. Au coated nano capillary membrane (NCAM) further enhanced fluorescent signal twelve times due to the increased surface area provided by the nanopore walls of the membrane and increased surface roughness. This NCAM sensor also allowed regenerations by

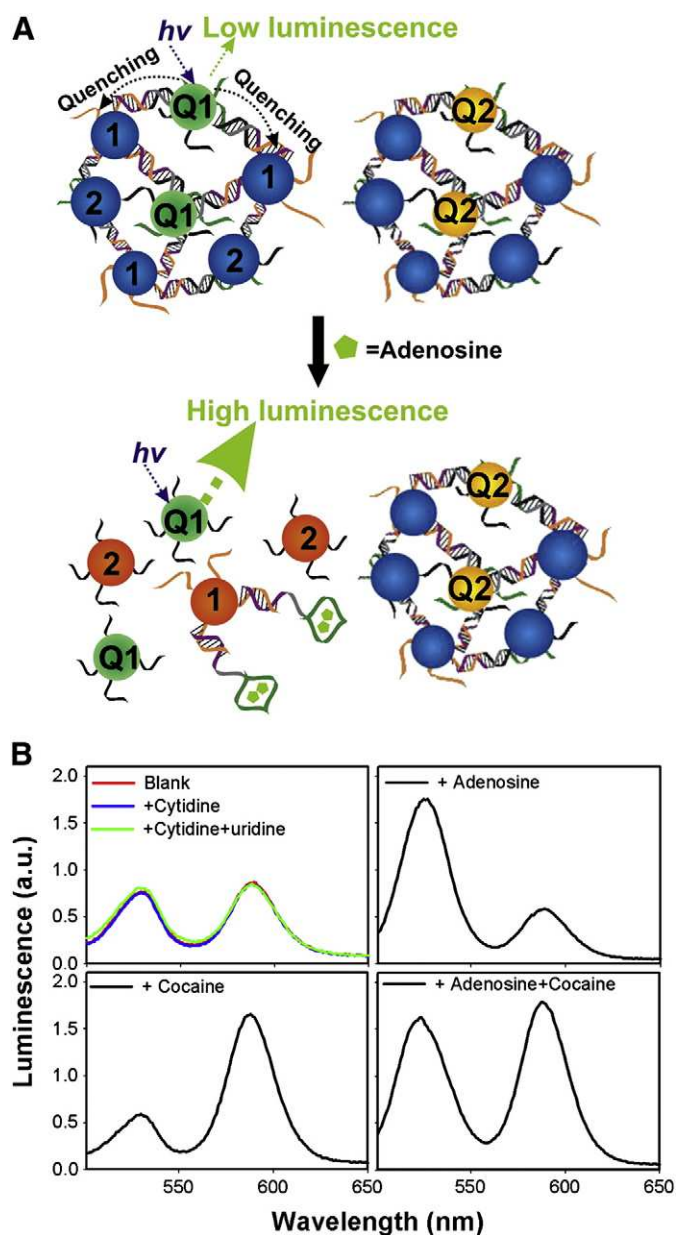


Fig. 3. Multiplexed detection of analytes using QDs. (A) Aptamer conjugated QD-AuNP aggregates are formed with two different QDs emitting at different wavelengths (QD1 and QD2). QDs in aggregated states are quenched due to the energy transfer from each QD to AuNPs. The fluorescence of QD1, however, is recovered after introduction of a target analyte (e.g. adenosine) as analyte-aptamer interaction causes disassembly of QD-AuNP aggregates. (B) Introduction of adenosine and/or cocaine increases the fluorescence of QD1 (emission at 525 nm) and/or QD2 (emission at 585 nm), respectively. On the other hand, the control analytes (cytidine and uridine) do not induce fluorescence signal increase from QDs. Reproduced with permission of ref. 96, copyright of American Chemical Society.

rehybridization of fresh DNA strand and storage up to 30 days without significant loss of activity [101]. Solid supported fluorescent sensors can also incorporate ratiometric fluorescence internal controls which allow to standardize the number of molecules immobilized on the surface and to perform real time detection of target molecules [102]. In addition, aptamers or DNAzymes can be incorporated into microfluidic devices which have advantages of generating low waste volume output, consuming less amount of detecting material, and having facile regeneration capabilities [103,104]. The Pb^{2+} DNAzyme integrated NCAM based microfluidic device consumed 4.2 picoliter of DNAzyme for each detection with detection limit of 11 nM [105]. Finally, by changing

the design of the microfluidic device, multiple detections could be made in a single cycle with minimal detection variation [106].

3.2. Colorimetric sensor

As fluorescence based sensors rely on the emission of the fluorophores or quantum dots after excitation at a certain wavelength, it is necessary to have analytical equipments such as fluorimeter or fluorescence microscope for detection. For real-time and on-site use, however, it is more convenient if detection can be carried out without any equipment. Colorimetric detection provides an advantage in this regard, since it allows detection to be made by naked eye [16,107–111]. Novel metallic nanoparticles, such as gold or silver nanoparticles, are ideal materials that allow colorimetric detection [112]. AuNPs have very high extinction coefficient, making their colors distinguishable without any instrument at only a few nanomolar concentration. Dispersed AuNPs smaller than 100 nm in solution originally have reddish color. When AuNPs aggregate, their color changes from red to blue due to a shift of their surface plasmon resonance to a higher wavelength [113].

There are two different methods for designing AuNP-DNA based colorimetric sensors. The labeled method is based on directed assembly (or disassembly) of AuNPs caused by analyte-specific binding, cleavage, or conformational change of the DNA molecules [8,16,113]. Two batches of AuNPs, each functionalized with a different DNA strand via Au-thiol chemistry, are mixed together, resulting in dispersed AuNPs due to the strong negative charge of DNA modified on AuNP surface. In the presence of a bridging DNA strand, which is complementary to the two different DNA strands functionalized on AuNP surface, AuNPs aggregate with their color changing from red to blue. The label-free method takes advantage of different adsorption properties of single stranded (ss) DNA and double stranded (ds) DNA onto citrate modified AuNPs [114]. Citrate modified AuNPs are naturally unstable in the presence of NaCl and can easily aggregate. Since ssDNA is flexible and can partially uncoil its strand, it can be readily adsorbed on AuNPs surface resulting in the enhancement of electrostatic repulsion between AuNPs, which in turn stabilizes AuNPs even in the presence of NaCl. On the other hand, as dsDNA is stiff and is covered with negatively charged phosphate backbone, it will repel the negatively charged citrate modified AuNPs, resulting in aggregation of AuNPs.

Both labeled and label-free methods have been used for colorimetric detection of DNA [12–14], RNA [15], and various metal ions [16–19,24,26,28,29] when DNA or DNAzymes are incorporated into the systems. Recently, a systematic comparison of the two methods using the same DNAzyme has been reported, providing a practical guidance in how to choose between the two methods [115].

Aptamers can also be incorporated into both systems for colorimetric detection. In 2006, we reported colorimetric sensors specific for adenosine and cocaine using the labeled method [111]. In the absence of target molecule, the aptamer-modified AuNPs remained aggregated, appearing blue; in the presence of the target, the AuNPs disassembled, resulting in red color (see Fig. 4A). The color change occurred in less than 1 minute. The labeled method has been used for detection of other small molecules and proteins [108,110]. Dong, Wang, and coworkers demonstrated that labeled aptamer-AuNP colorimetric system could be used for detection of thrombin with extremely high sensitivity (with detection limit of 14 fM) using dot-blot arrays [116]. Furthermore, our group reported multiplex colorimetric sensing system based on smart AuNPs responsive to multiple stimuli with controlled cooperativity [117].

Fan and co-workers reported colorimetric detection of ATP using unmodified AuNPs and ATP aptamers based on label-free method (see Fig. 4B) [118]. The AuNPs aggregated displaying a blue color in the absence of ATP but remained dispersed displaying a red color in its presence. The sensor had high selectivity and a detection limit of

0.6 μM . This label-free aptamer based colorimetric detection method was also used for other analytes such as potassium [19], cocaine [118], thrombin [119].

Li and coworkers reported a detection method based on a combination of labeled and label-free methods (see Fig. 4C). They first chemically functionalized short DNA strands on AuNPs and hybridized those strands with long partially complementary aptamer strands. The long aptamer DNA provides additional negative charge to AuNPs, enhancing their stability at a certain concentration of NaCl. Introduction of target molecules caused interaction between the target molecule and the aptamer strand, resulting in the release of the aptamer strand from AuNP. The targeted release of aptamers decreased the number of negatively charged DNA strands on AuNPs, lowering the stability of AuNPs. This turn-on colorimetric sensor,

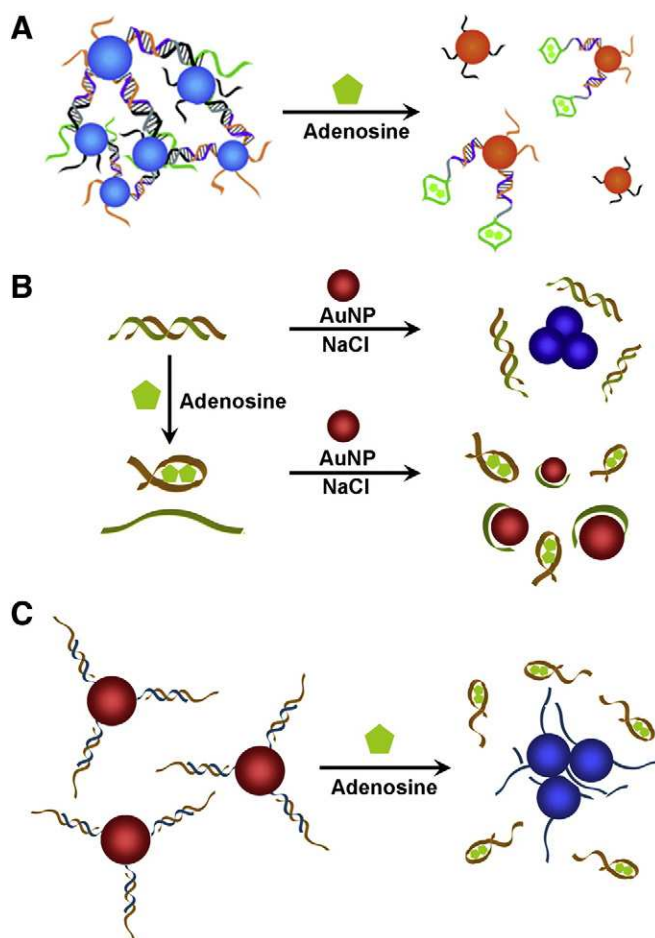


Fig. 4. Aptamer-AuNP based colorimetric sensors. (A) AuNPs functionalized with oligonucleotides have been used for labeled sensors. AuNPs are initially aggregated with adenosine specific aptamer strands. The introduction of adenosine stimulates disassembly of AuNPs, changing the color of AuNPs from blue to red. (B) The interaction between unmodified AuNPs and unlabeled ssDNA strand has been used to make label-free sensors. In the presence of adenosine, dsDNA containing an adenosine aptamer strand (brown) releases ssDNA (green) which can be adsorbed onto unmodified AuNPs. AuNPs remain dispersed with red color even in the presence of NaCl due to enhanced stability of AuNPs provided by ssDNA. In the absence of adenosine, dsDNA with adenosine aptamer strand stays hybridized. The electrostatic repulsion between dsDNA and AuNP as well as the stiffness of dsDNA make them ineffective in preventing NaCl induced charge screening on AuNP surface, causing aggregation of AuNPs with color change from red to blue. (C) Both labeled and label-free method has been combined to make an adenosine sensor. AuNPs are chemically functionalized with DNA (blue) and hybridized with adenosine aptamer strand (brown). In the presence of adenosine, aptamer strands are released from AuNPs, decreasing the stability of AuNPs. The resulting aggregation of AuNPs causes their color change from red to blue.

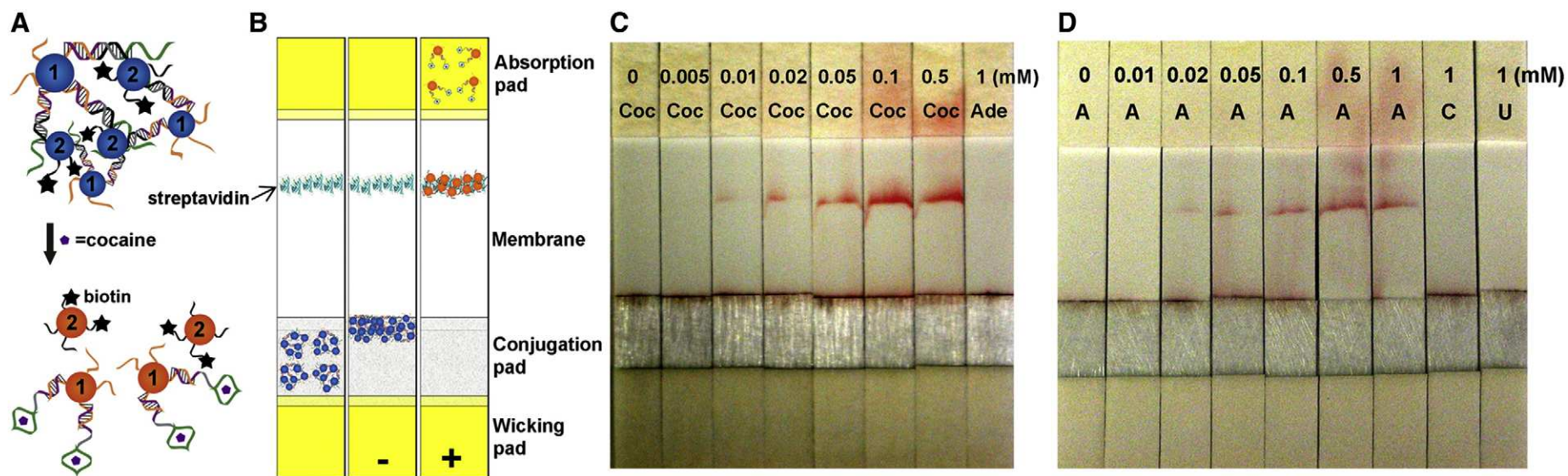


Fig. 5. Simple dip stick tests based on labeled AuNP-aptamer lateral flow device. (A) Schematic illustration of cocaine aptamer induced AuNP disassembly and incorporation of such system onto the lateral flow device. Aggregates of AuNPs functionalized with both cocaine aptamer and biotin are disassembled in the presence of cocaine due to the binding between aptamer and cocaine. (B) The structure of lateral-flow device; the device is composed of a wicking pad, a glass fiber conjugation pad, a membrane, and an absorption pad. Biotin labeled AuNP aggregates containing cocaine aptamers are dropped onto the conjugation pad and the wicking pad is dipped into a sample solution with cocaine. The solution flows along the conjugation pad, rehydrates and induces disassembly of AuNP aggregates. (C) The dipstick test for cocaine. In the presence of cocaine, disassembled AuNPs with biotin are captured onto streptavidin immobilized on the membrane, producing a red line. (D) The lateral flow device can be generally applied with other aptamers, such as adenosine aptamers, for adenosine sensing. Reproduced with permission of ref. 124. Copyright Wiley-VCH Verlag GmbH & Co. KGaA.

changing its color from red to blue in the presence of analytes, had high selectivity and a detection limit of $10\ \mu\text{M}$ [120,121].

While AuNP based colorimetric sensors provide a way to carry out detection without the need for analytical equipment, it still requires careful handling of micro-liter scale solutions, which is not trivial for most users, especially those at home. Therefore it is preferred to develop more user-friendly diagnostic kits, such as dip-stick test. Since the discovery of antibodies, there have been a number of reports converting antibody based detection onto the lateral flow devices [122] and many of the kits, such as home pregnancy test, are commercially available. In contrast, few reports demonstrating nucleic acid based lateral flow devices have appeared in the literature. For example, Ioannou and Christopoulos *et al.* used DNA functionalized AuNPs to analyze the hybridization of DNA on lateral flow devices [123]. We developed a lateral flow device based on labeled AuNP and aptamer system (see Fig. 5) [124]. The lateral-flow device is composed of four components, a wicking pad, a glass fiber conjugation pad, a membrane, and an absorption pad. Biotin labeled AuNP aggregates containing cocaine aptamers were dropped onto the conjugation pad, and streptavidin was immobilized on a specific part of the membrane to capture biotinylated AuNPs. When the wicking pad of lateral flow device was dipped into a sample solution containing cocaine, the solution flowed along the conjugation pad, rehydrated and induced disassembly of AuNP aggregates. Since biotin was labeled on disassembled AuNPs, the AuNPs could be captured on the part of the membrane with streptavidin forming a red line. When the concentration of cocaine in the solution increased, the intensity of the red line formed on the membrane became higher and the estimated detection limit was $\sim 10\ \mu\text{M}$. In addition to the simplicity in testing without the need of precise solution transfer, the method was also more sensitive than many solution-based tests, due to the integration of binding, separation, and detection on a simple test paper-like platform. Red lines did not appear with control analytes, showing good selectivity of the lateral flow device. In the process, we discovered a new design of the lateral flow device taking advantage of the physical size difference of nanoparticles in various assembly states, which provides a critical control for assessing the performance of the device. The lateral flow device could also be applied to adenosine by using the aptamer specific for the molecule. Since aptamers for a broad range of molecules have been obtained, this method is general enough to be adapted to develop “dip-stick” tests for any analyte for which an aptamer can be obtained. In addition, the lateral flow device also performed well in human blood serum, showing its compatibility with biological samples. Recently, Pelton, Li and coworkers reported that aptamers can be immobilized on cellulose surface as a potential platform for detection of medically relevant biomolecules [125], making it possible for medical diagnostics.

3.3. Surface Enhanced Raman Scattering (SERS)

Raman scattering is an optical methodology utilizing the inelastic scattering of a photon. It has several advantages over fluorescence as it can provide structural information of a molecule, does not suffer from photobleaching, and has a much narrower spectral width ($<1\ \text{nm}$, half widths), which is advantageous for multiplexed detection. As Raman cross section is inherently weak and inefficient ($10^{-30}\ \text{cm}^2$ per molecule vs $10^{-16}\ \text{cm}^2$ per molecule for fluorescence), there has been limitation in utilizing Raman scattering as a tool for highly sensitive detection [126]. In 1974, however, it was reported that Raman scattering can be significantly amplified when molecules are adsorbed on metal surface, due to the extremely high electromagnetic fields produced on hot spots generated on the surface of metals [127]; this phenomenon is called Surface Enhanced Raman Scattering (SERS). SERS becomes more efficient on rough metal surfaces or on metal nanoparticle aggregates due to excitation of localized surface plasmon [128].

In 1997, Nie and Kneipp independently demonstrated the possibility of detecting single molecules based on SERS [126,129]. Using Ag colloidal nanoparticles (AgNPs), Nie reported that the enhancement factor of 10^{14} – 10^{15} and the cross section of $10^{-16}\ \text{cm}^2$ per molecule can be obtained which are comparable to fluorescence [126]. Taking advantage of AuNPs coated with Ag as surface enhancement promoter with Raman reporters, highly sensitive and selective multiplexed detection of DNA was reported by Mirkin and co-workers [130]. Halas *et al.* demonstrated that SERS spectra can be obtained from DNA without any Raman reporters although the obtained spectra were mostly identical for several model DNA strands, which are dominated by adenine [131].

The detection of proteins or small molecules using SERS has also been reported using DNA aptamers. Wang, Dong, and co-workers used aptamers to detect α -thrombin by SERS (See Fig. 6A) [132]. Thrombin has two binding sites specific for two different DNA aptamers with high affinity [61,133]. A substrate functionalized with aptamer 1 was first treated with thrombin and then incubated in AuNPs functionalized with aptamer 2 and SERS reporter. As a result, AuNPs were attached to the surface through aptamer-thrombin interaction. After Ag NP deposition, the NPs became larger, resulting in highly enhanced Raman scattering and thus highly sensitive signal. The detection limit was reported as $0.5\ \text{nM}$ and the system had high selectivity over both β - and γ - thrombins. Jiang, Yu, and co-workers demonstrated a different methodology to detect adenosine using aptamers [134]. They used a dsDNA composed of both an adenosine aptamer strand and a partially complementary short DNA strand with a Raman reporter (Fig. 6B). In the presence of adenosine, the aptamer strand underwent structure switching, resulting in denaturation of dsDNA. Therefore the Raman probe labeled oligonucleotide was released in solution. The

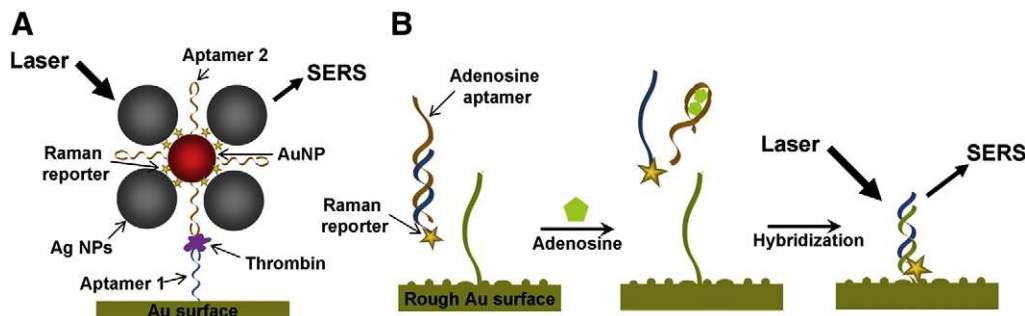


Fig. 6. Detection of thrombin by SERS. (A) SERS based thrombin sensing. Thrombin is first conjugated on thrombin aptamer 1 (blue) immobilized on Au surface. AuNPs with both Raman reporters and thrombin aptamer 2 (brown) is then attached on the thrombin pre-conjugated on aptamer 1 via the interaction between thrombin and aptamer 2. After Ag NP deposition, amplified Raman signal can be obtained. (B) SERS based adenosine sensing. In the presence of adenosine, structure switching of adenosine aptamer strand (brown) of dsDNA causes the release of the short ssDNA strand (blue) with Raman reporter. The ssDNA with a Raman reporter is hybridized to the ssDNA (green) immobilized on rough Au surface, inducing amplified Raman scattering.

released strand later hybridized to the complementary DNA which was previously immobilized on a rough gold surface. This chain of processes resulted in an enhanced Raman signal.

SERS can not only be used *in vitro*, but also be used *in vivo* for tumor targeting. Nie and co-workers recently reported that polyethylene glycol (PEG) modified 80 nm AuNPs which incorporated both Raman reporter and antibody modification could be used as SERS nanoparticles for targeted imaging of tumors [135]. Gambhir and coworkers also demonstrated non-invasive whole body Raman imaging and *in vivo* tumor imaging using SERS nanoparticles and single wall CNTs [136].

3.4. Magnetic Resonance Imaging (MRI)

While fluorescence and SERS have been used in imaging *in vivo*, the penetrating length is limited. MRI, on the other hand, does not have this limitation and allows 3D imaging of the whole human body with clinically available instruments all over the world. Currently, MRI is mostly effective only when the difference in tissues, such as between a solid tumor and a regular tissue, is large, with little molecular information. For future MRI imaging, it is preferred to use MRI to detect molecular markers of the tumor before the tumor develops. To achieve this goal, new smart MRI contrast agents responsive to small molecular markers are required.

Superparamagnetic iron oxide nanoparticles are a new class of MRI contrast agents, catching the attention of many researchers due to their advantageous properties over gadolinium-based contrast agents.

It has been shown in literature that superparamagnetic iron oxide nanoparticles can be coupled to biological molecules including thiol modified oligonucleotides for active targeting. Weissleder and co-workers demonstrated that cross-linked dextran iron oxide (CLIO) nanoparticles can be functionalized with DNA and this material can be used to detect sequence specific oligonucleotide by MRI [137,138]. CLIO nanoparticles change the spin-spin relaxation time (T₂) of surrounding water protons. In addition, researchers observed that these nanoparticles become more efficient in decreasing T₂ when assembled into bigger clusters. This observation has been confirmed by many different assembly and disassembly systems [137,139–141]. Based on this principle it is possible to distinguish assembled and disassembled states of the nanoparticles and therefore detect the analytes which results in such transition.

Recently we have developed a method for detecting chemical and biological molecules using aptamer functionalized CLIO nanoparticles. Adenosine aptamer could induce disassembly of CLIO nanoparticle aggregates in the presence of adenosine which was observed by MRI with an increase in the brightness of T₂-weighted MR images (Fig. 7A) [142]. The control analytes without any activity on adenosine aptamer did not show any change in MR images. The CLIO nanoparticles functionalized with mutated aptamer (which does not interact with adenosine) did not show any change in MR image, either. In order to demonstrate the response on proteins instead of small molecules and observe the change in contrast, we have tested the system for thrombin [143]. The CLIO nanoparticles functionalized with thrombin aptamers assembled in the presence of thrombin, which could be

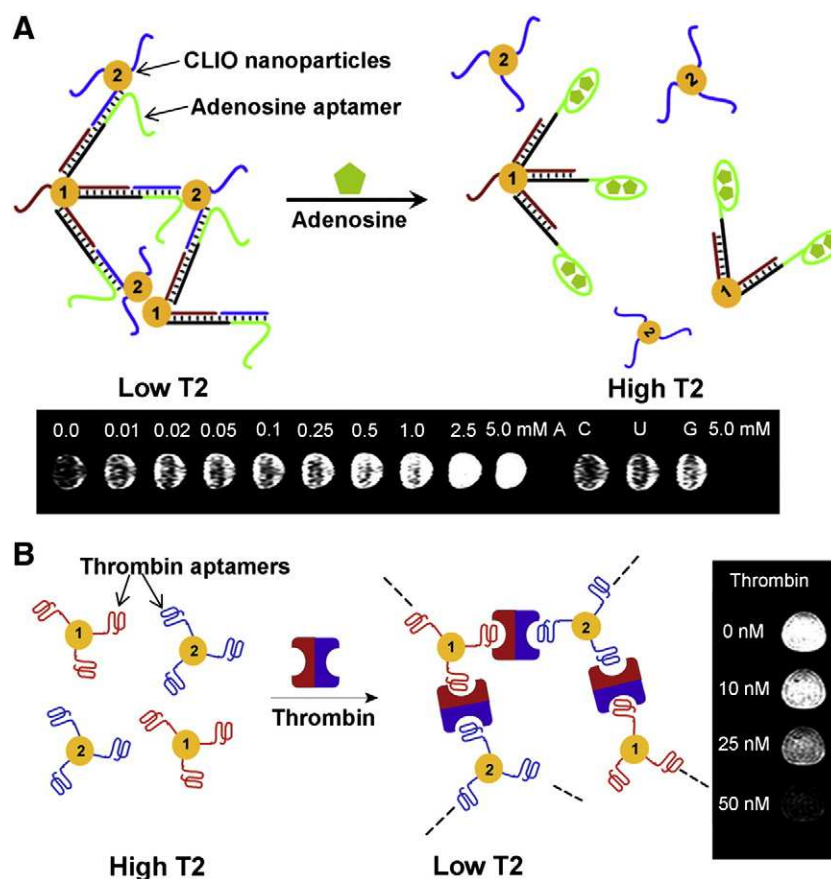


Fig. 7. Smart MRI contrast agents based on aptamer functionalized CLIO nanoparticles. (A) MRI based small molecule sensing. Aggregates of CLIO nanoparticles were formed by hybridization of two strands of DNA (brown and blue) functionalized on CLIO nanoparticles with the complementary DNA containing adenosine aptamer (green). The CLIO nanoparticle aggregates disassembled in the presence of adenosine, due to the structure switching of aptamers after binding to adenosine, which could be observed by the increase of T₂ value and brighter MR images. (B) MRI based protein sensing. The CLIO nanoparticles functionalized with two different thrombin aptamers (red and blue) assemble in the presence of thrombin, which could be observed by the decrease of T₂ value and darker MR images. Fig. 7A is reproduced with permission from ref. 142, copyright of Wiley-VCH Verlag GmbH & Co. KGaA. Fig. 7B is reproduced with permission from ref. 143, copyright of the American Chemical Society.

observed by the decrease of brightness in T2 weighted MR images (Fig. 7B). Such an aptamer functionalized superparamagnetic iron oxide nanoparticles are suitable for non-invasive *in vivo* imaging of small molecular markers, making early diagnostics and treatment of diseases possible before the progression of the disease to late stages.

3.5. Electrochemical detection

Most of the optical detection methods require highly transparent sample solution as the color of the solution might interfere with the optical signal generated from sensors. For biomedical diagnostics on biological samples, such as blood, it is necessary to find an alternative method. Electrochemical detection provides another important route for non-transparent samples since the signal transduction can be accomplished by non-optical means [144].

Electrochemical detection is carried out by measuring changes in redox states of redox active probes. As the labels can be easily attached to DNA, electrochemical method has been used extensively to detect DNA [145–147]. There are several different strategies reported for electrochemical detection of analytes using aptamers. One takes advantage of the redox state change induced by catalysts immobilized on DNA aptamers. Since thrombin has two binding sites for two different aptamer strands [61,133], it is possible to immobilize thrombin on Au electrode with one aptamer and bind a catalyst which is tethered on the other aptamer. An example of using platinum nanoparticle (Pt NP) as a catalyst label is shown in Fig. 8A [148]. The thrombin aptamer 1 was first immobilized on Au electrode and allowed to bind to thrombin. The Pt NP tethered with thrombin aptamer 2 was then introduced to form thrombin-aptamer-Pt NP sandwich complex. The Pt NPs catalyzed H_2O_2 reduction to H_2O , resulting in cathodic currents which enabled

thrombin detection with a detection limit as low as 1 nM. Similar methods using biocatalysts were also reported [149,150] but Pt NPs showed higher sensitivity, probably due to the effectiveness of the Pt NP catalyst. It is also possible to carry out electrochemical sensing based on the conformation change of redox reporter labeled aptamers immobilized on Au electrode for detection of small molecules [151–153] and protein [154–157].

The electrochemical sensors mentioned above are highly sensitive. To extend the methodology to multiplexed detection, Wang and co-workers used protein labeled QDs with different chemical compositions [158]. In this work, they immobilized two different aptamers on a Au surface which were specific for thrombin and lysozyme, and then introduced both thrombin and lysozyme labeled with CdS and PbS QDs, respectively (see Fig. 8B). The protein analytes (either thrombin or lysozyme) were added afterwards, replacing corresponding QD labeled proteins on the surface with sample analytes. By monitoring the remaining QDs via electrochemical stripping detection, it was possible to identify the proteins in the sample solution. Using this method, sub-picomolar detection limit was achieved with high selectivity.

Willner and co-workers demonstrated label-free detection of adenosine mono-phosphate (AMP) using ion-selective field effect transistors (ISFET) [159]. In this work, they immobilized adenosine selective aptamers on Al_2O_3 gate surface and hybridized aptamers with short complementary DNA strands (see Fig. 9A). The addition of AMP induced structure switching of aptamer strand, causing release of short partially complementary DNA strand. This released complementary strand altered the charge associated on the gate, changing the source to drain current. Gate-to-source potential (V_{gs}) of the device increased as the concentration of AMP became higher and the detection limit of the device was 50 μ M.

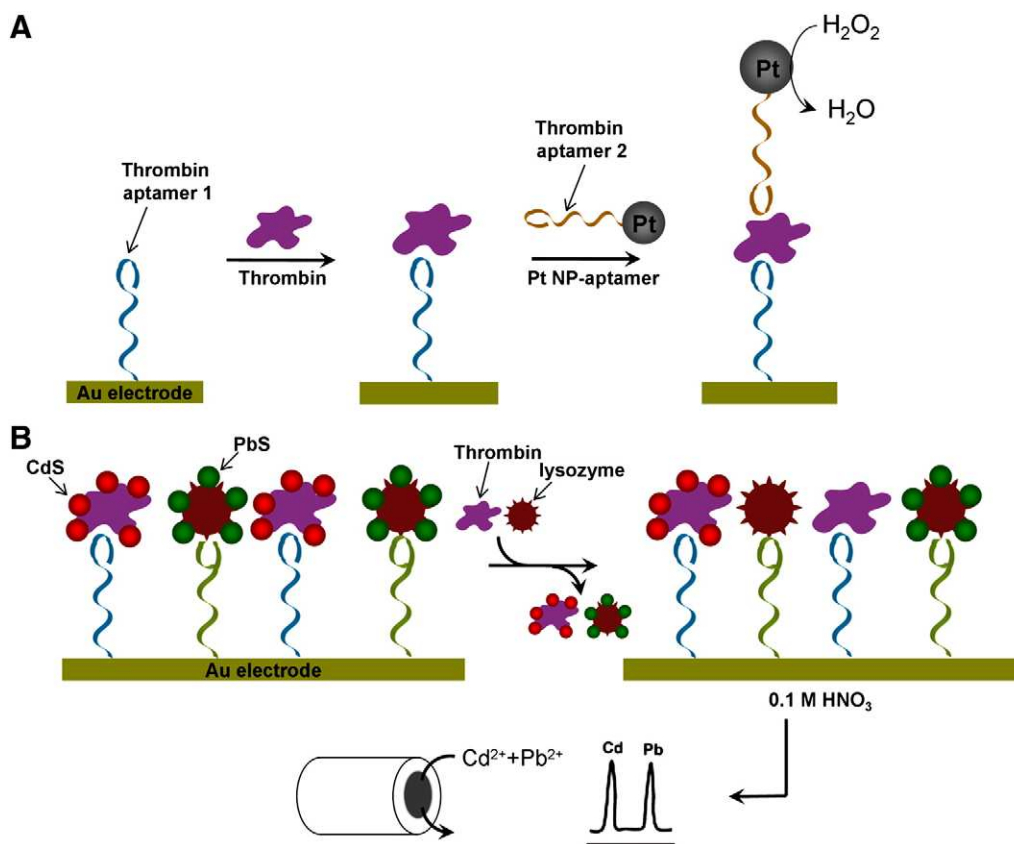


Fig. 8. Electrochemistry based multiplexed detection of proteins. (A) Electrochemical sensor for the detection of thrombin, based on the redox change induced by the Pt NPs immobilized on DNA thrombin aptamers. The Pt NPs catalyze H_2O_2 reduction to H_2O , resulting in cathodic currents, enabling thrombin detection. (B) Multiplexed detection of thrombin and lysozyme. Aptamers for both proteins are immobilized on Au surface and CdS labeled thrombin and PbS labeled lysozyme are then bound on the aptamers, respectively. In the presence of protein analyte, the protein replaces corresponding QD labeled protein on the surface. The amount of proteins in sample solution can be identified by monitoring the remaining QD by electrochemical stripping detection. Reproduced with permission of ref. 158, copyright of American Chemical Society.

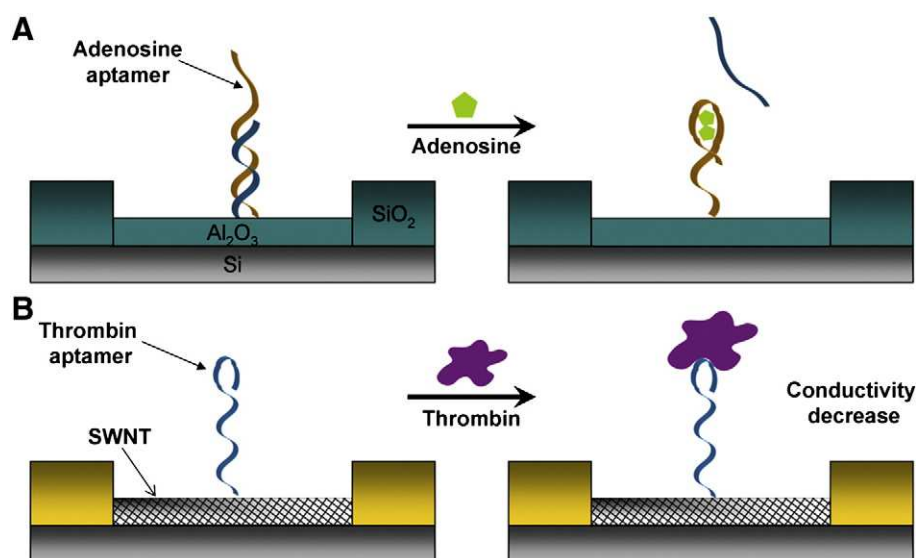


Fig. 9. Detection of chemical and biological molecules based on aptamer conjugated electronic devices. (A) Label-free detection of AMP using ion-selective field effect transistors. Adenosine aptamer (brown) is immobilized onto Al₂O₃ gate surface and hybridized with a partially complementary DNA strand (blue). In the presence of adenosine complementary strand is released, inducing change of the source-to-drain current. (B) CNT based field effect transistor sensor for the detection of thrombin. Thrombin aptamers (blue) are immobilized on CNT field effect transistor. The introduction of thrombin decreases the conductance of field effect transistor.

Carbon nanotube (CNT) is an interesting nanomaterial with either metallic or semiconductive properties based on its structure, such as chirality or diameter [160–162]. As CNTs have the size (c.a. 1–2 nm diameter) comparable to most biomolecules as well as interesting physical and chemical properties, there has been intense investigation to conjugate CNTs with biomolecules to form a hybrid system. It has been reported that DNA can either noncovalently bind to the external part of CNT walls after electrical, thermal, or sonication treatment [163], or covalently bind to the carboxyl groups introduced on the CNT sidewalls after oxidization treatment [164–167]. Bifunctional reagents can also be used to introduce functional groups on DNA [168,169].

CNTs can be used as a conducting channel in field effect transistors (FET). As the interaction between biomolecules and CNTs can modulate the conductivity of the FET, detection can be accomplished based on the change of electrical signal. Detection of DNA has been demonstrated using CNT based FETs.

Lee and co-workers were the first to report the detection of adenosine using aptamer conjugated single wall carbon nanotube (SWNT) field effect transistor (FET) [170]. In their work, they modified the surface of CNT with CDI-Tween via hydrophobic interaction [169] and covalently attached 3'-amine modified thrombin aptamer DNA on it (Fig. 9B). As soon as thrombin was dropped on the surface of the device, the conductance of the device decreased abruptly and soon saturated, which might be attributed to the screening of negative charges of DNA aptamers or positive charge induced by the introduction of thrombin. The detection limit was as low as 10 nM and the device had selectivity over the protein elastase, which is another serine protease with molecular weight and isoelectric point similar to thrombin.

It is reported that aptamers work much more efficiently than antibodies in CNT FET sensors. Tamiya and co-workers compared the performance of IgE specific aptamer and monoclonal antibody for the detection of immunoglobulin E (IgE) on CNT FET system [171]. Even though they could observe IgE induced conductance decrease for both aptamers and antibodies systems, the aptamer system worked much more effectively than antibody with the detection limit of 250 pM. The reason is mainly due to the small size of aptamers (1–2 nm) compared to antibodies (~10 nm) that allows target protein to bind to CNT within the electrical double layer in physiological conditions, making the detection possible. This method is versatile and its application for

the detection of microorganism was also reported using *Escherichia coli* (*E. coli*) as an example [172]. By employing RNA based *E. coli* aptamers, the study demonstrated the detection of *E. Coli* in less than 20 minutes with selectivity over salmonella.

4. Targeted drug delivery in cells using aptamer-nanomaterial conjugates

In addition to molecular diagnostics and imaging discussed above, the aptamer-nanomaterial conjugates have also been applied in targeted drug delivery, making it possible for therapeutic applications. Farokhzad, Langer, and co-workers reported a smart QD-aptamer conjugate which serves both as a fluorescence imager and a drug delivery vehicle (See Fig. 10) [173]. The QD-aptamer conjugate was composed of 3 components; QD, prostate cancer (PCa) cell specific RNA aptamer, and doxorubicin (Dox) which is commonly used anthracycline drug with fluorescence when intercalated into CG pairs of double stranded oligonucleotide. Aptamers were conjugated to QDs and Dox was intercalated to the aptamer strand [174], forming QD-aptamer-Dox system. The QD-aptamer-Dox system was initially 'off' since the fluorescence of QD was transferred to Dox and the fluorescence of Dox was also quenched by the dsRNA aptamer, both via energy transfer between the two components. Once injected into cancer cell, the QD-aptamer-Dox system gradually released Dox, caused by the binding of target molecule onto RNA aptamer. This Dox release recovered the fluorescence of the QD. Therefore the QD-aptamer-Dox system not only allows targeted drug delivery, but also enables monitoring drug release and imaging target cells. A similar methodology has also been applied to superparamagnetic iron oxide nanoparticles for prostate cancer imaging and therapy [175].

Lippard, Farokhzad, Langer, and coworkers demonstrated aptamer functionalized Pt(IV) prodrug-PLGA-PEG nanoparticles for targeted delivery of cisplatin to prostate cancer cells [176]. Prostate-specific membrane antigen (PSMA) targeting aptamer functionalized controlled release block-copolymer nanoparticles could target and be taken up by human prostate PSMA overexpressing LNCaP epithelial cells with high efficiency, but not to PC3 cancer cells. Furthermore, the aptamer-Pt(IV) encapsulated NPs were much more effective than the free cisplatin or nontargeted nanoparticles against the LNCaP cells for drug delivery.

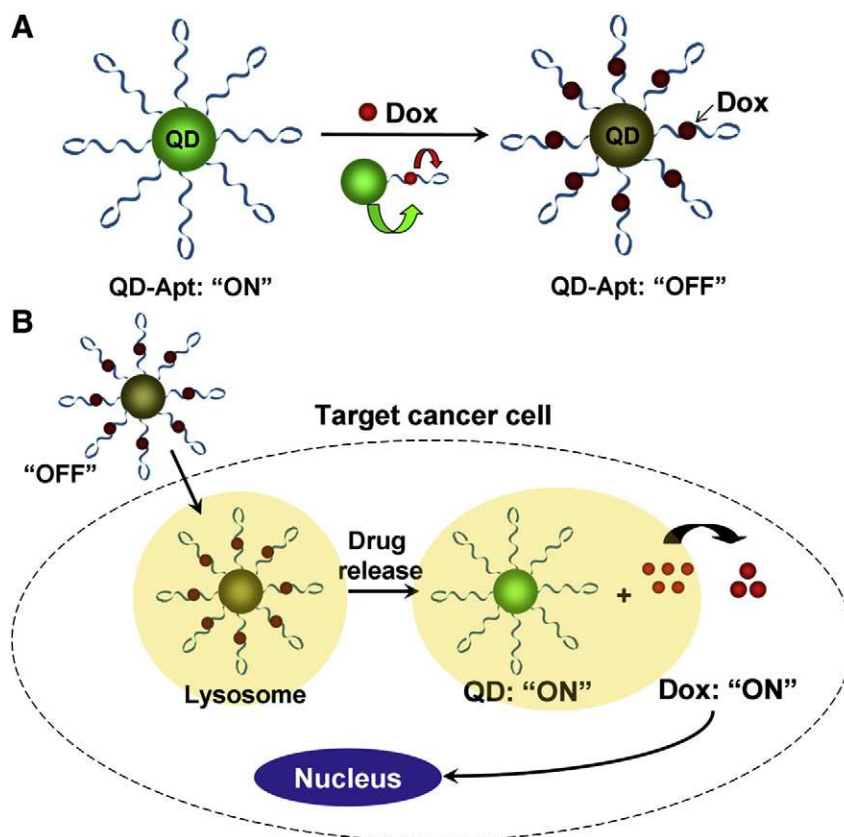


Fig. 10. QD-aptamer conjugate serving as both a fluorescence imaging agent and a drug delivery vehicle. (A) The conjugate is composed of QD, RNA aptamer, and doxorubicin. (Dox). QD-aptamer-Dox system is initially 'off' as the fluorescence of QD is transferred to Dox and the fluorescence of Dox is quenched by the dsRNA aptamer, both by energy transfer. (B) Once placed inside cancer cells, Dox is gradually released from QD-aptamer-Dox system and fluorescence of the QD is recovered.

In addition, Langer and coworkers were the first to demonstrate aptamer based drug delivery *in vivo* [50]. They showed that docetaxel (Dtxl) containing di-block copolymer nanoparticles functionalized with RNA aptamers could significantly decrease tumor with reduced toxicity compared to Dtxl alone.

Tan and coworkers demonstrated an advantage of using multiple aptamer-conjugated nanorods as a multivalent probe for targeted cancer cell imaging [177]. Many targeting molecules, such as antibodies or aptamers, have moderate binding affinities which prevent efficient targeted cell imaging. By using Au-Ag nanorods conjugated to up to 80 fluorophore labeled aptamers, 300 fold increased fluorescence signal and 26 times higher binding affinity was obtained compared to individual aptamer strand. This methodology enables to target and image cells with low density binding sites or targeting molecules with relatively low binding affinities.

Targeted delivery of DNA or RNA molecules into cells is among one of potent therapeutic strategies for genetic diseases such as cancer. However, practical application of DNA or RNA delivery in cellular environment is difficult as those molecules can be generally degraded or digested by nucleases or enzymes. It is reported that SWNTs can be used as a protective cargo for delivery of ssDNA into cells [178]. The work from Tan and coworkers demonstrated the stability of delivered DNA against nuclease and enzymes *in vitro*. In addition, it has been shown that specific mRNA targeting DNA could be delivered efficiently into cells in DNA/SWNT-complexed form and the delivered DNA was active. This work demonstrated the possibility of using SWNTs as DNA/RNA delivery molecules for diagnostic and therapeutic applications. Furthermore, it has been reported recently that viral capsid carrier can also be functionalized to DNA aptamer and these can potentially be used for targeted drug delivery [179]. Capsids functionalized with 41 mer DNA aptamer specific for tyrosine

kinase receptor could successfully target and be uptaken by Jurkat T cells.

We, in collaboration with Wong and Cheng groups, recently reported cell specific drug delivery based on aptamer functionalized liposomes [180]. Liposome is, by far, the most widespread system used for drug delivery. By conjugating aptamers specific to nucleolin, which is overexpressed on cancer cell plasma membrane, along with cisplatin encapsulated inside liposomes, we demonstrated targeted delivery of cisplatin onto nucleolin overexpressing breast cancer MCF-7 cells but not to control prostate cancer cells (LNCaP cell). Furthermore, the extent of drug release can be controlled by using a complementary DNA of the aptamer as an antidote so that the drug effects can be readily controlled.

5. Conclusion

By summarizing recent progress in developing methods of integrating aptamers with a diverse number of nanomaterials, we have shown that these novel bionanomaterials can be used as highly sensitive and selective diagnostic agents and targeted drug delivery agents. As each nanomaterial has different optical, electrochemical, magnetic, and mechanical properties, medical diagnostic and drug delivery agents with diverse characteristics can be used for different biomedical applications, making them an excellent alternative to antibody-based medical applications.

Most of the agents introduced here, however, used available aptamers specific for molecules such as adenosine, cocaine and thrombin, as a proof of concept only, and most of tests have been carried out *in vitro*. To realize the full potentials of such bionanomaterials systems, one needs to carry out more selections for aptamers that are specific for more clinically relevant targets such as cancer markers and viruses, and demonstrate the effectiveness of the systems

in vivo. Given progresses made in the field in such a short period of time, applications of these aptamer-nanomaterial based sensors in clinical applications will be realized in the near future.

Acknowledgements

We thank the US National Institute of Health (ES016865) and the National Science Foundation (DMR-0117792, CTS-0120978 and DMI-0328162) for financial support.

References

- [1] C.M. Lieber, *Nanoscale Science and Technology: Building a Big Future from Small Things*, MRS Bull. 28 (2003) 486.
- [2] M.-C. Daniel, D. Astruc, *Gold Nanoparticles: Assembly, Supramolecular Chemistry, Quantum-Size-Related Properties, and Applications toward Biology, Catalysis, and Nanotechnology*, Chem. Rev. 104 (2004) 293.
- [3] J. Li, Y. Lu, A Highly Sensitive and Selective Catalytic DNA Biosensor for Lead Ions, *J. Am. Chem. Soc.* 122 (2000) 10466.
- [4] S.D. Jhaveri, R. Kirby, R. Conrad, E.J. Maglott, M. Bowser, R.T. Kennedy, G. Glick, A.D. Ellington, *Designed Signaling Aptamers That Transduce Molecular Recognition to Changes in Fluorescence Intensity*, *J. Am. Chem. Soc.* 122 (2000) 2469.
- [5] Y. Lu, *New Transition Metal-Dependent DNAszymes as Efficient Endonucleases and as Selective Metal Biosensors*, Chem. Eur. J. 8 (2002) 4588.
- [6] N.K. Navani, Y. Li, *Nucleic Acid Aptamers and Enzymes as Sensors*, Curr. Opin. Chem. Biol. 10 (2006) 272.
- [7] Y. Lu, J. Liu, *Functional DNA Nanotechnology: Emerging Applications of DNAszymes and Aptamers*, Curr. Opin. Biotechnol. 17 (2006) 580.
- [8] Y. Lu, J. Liu, *Smart Nanomaterials Inspired by Biology: Dynamic Assembly of Error-Free Nanomaterials in Response to Multiple Chemical and Biological Stimuli*, Acc. Chem. Res. 40 (2007) 315.
- [9] J. Liu, Z. Cao, Y. Lu, *Functional Nucleic Acid Sensors*, Chem. Rev. 109 (2009) 1948.
- [10] Z. Wang, Y. Lu, *Functional DNA Directed Assembly of Nanomaterials for Biosensing*, *J. Mater. Chem.* 19 (2009) 1788.
- [11] Y. Li, Y. Lu, *Functional Nucleic Acids for Sensing and Other Analytical Applications*, Springer, New York, NY, 2009.
- [12] R. Elghamian, J.J. Storhoff, R.C. Mucic, R.L. Letsinger, C.A. Mirkin, *Selective Colorimetric Detection of Polynucleotides Based on the Distance-Dependent Optical Properties of Gold Nanoparticles*, Science 277 (1997) 1078.
- [13] P. Hazarika, B. Ceyhan, C.M. Niemeyer, *Reversible Switching of DNA-Gold Nanoparticle Aggregation*, Angew. Chem. Int. Ed. 43 (2004) 6469.
- [14] H. Li, L. Rothberg, *Colorimetric Detection of DNA Sequences Based on Electrostatic Interactions with Unmodified Gold Nanoparticles*, PNAS 101 (2004) 14036.
- [15] H. Li, L. Rothberg, *Detection of Specific Sequences in RNA Using Differential Adsorption of Single-Stranded Oligonucleotides on Gold Nanoparticles*, Anal. Chem. 77 (2005) 6229.
- [16] J. Liu, Y. Lu, *A Colorimetric Lead Biosensor Using DNAszyme-Directed Assembly of Gold Nanoparticles*, *J. Am. Chem. Soc.* 125 (2003) 6642.
- [17] J. Liu, Y. Lu, *Stimuli-Responsive Disassembly of Nanoparticle Aggregates for Light-up Colorimetric Sensing*, *J. Am. Chem. Soc.* 127 (2005) 12677.
- [18] J.-S. Lee, M.S. Han, C.A. Mirkin, *Colorimetric Detection of Mercuric Ion (Hg²⁺) in Aqueous Media by DNA-Functionalized Gold Nanoparticles*, Angew. Chem. Int. Ed. 46 (2007) 4093.
- [19] L. Wang, X. Liu, X. Hu, S. Song, C. Fan, *Unmodified Gold Nanoparticles as a Colorimetric Probe for Potassium DNA Aptamers*, Chem. Commun. (2006) 3780.
- [20] J. Liu, A.K. Brown, X. Meng, D.M. Crokek, J.D. Istok, D.B. Watson, Y. Lu, *A Catalytic Beacon Sensor for Uranium with Parts-Per-Trillion Sensitivity and Millionfold Selectivity*, PNAS 104 (2007) 2056.
- [21] J. Liu, Y. Lu, *Rational Design Of "Turn-On" Allosteric DNAszyme Catalytic Beacons for Aqueous Mercury Ions with Ultrahigh Sensitivity and Selectivity*, Angew. Chem. Int. Ed. 46 (2007) 7587.
- [22] J. Liu, Y. Lu, *Colorimetric Cu²⁺ Detection with a Ligation DNAszyme and Nanoparticles*, Chem. Commun. (2007) 4872.
- [23] J. Liu, Y. Lu, *A DNAszyme Catalytic Beacon Sensor for Paramagnetic Cu²⁺ Ions in Aqueous Solution with High Sensitivity and Selectivity*, *J. Am. Chem. Soc.* 129 (2007) 9838.
- [24] C.W. Liu, Y.T. Hsieh, C.C. Huang, Z.H. Lin, H.T. Chang, *Detection of Mercury(II) Based on Hg²⁺-DNA Complexes Inducing the Aggregation of Gold Nanoparticles*, Chem. Commun. 19 (2008) 2242.
- [25] D.P. Wernet, J. Liu, P.W. Bohn, Y. Lu, *Functional-DNA-Based Nanoscale Materials and Devices for Sensing Trace Contaminants in Water*, MRS Bull. 33 (2008) 34.
- [26] D. Li, A. Wiekowska, I. Willner, *Optical Analysis of Hg²⁺ Ions by Oligonucleotide-Gold-Nanoparticle Hybrids and DNA-Based Machines*, Angew. Chem. Int. Ed. 47 (2008) 3927.
- [27] T.S. Dalavoy, D.P. Wernet, M. Gong, J.V. Sweedler, Y. Lu, B.R. Flachsbart, M.A. Shannon, P.W. Bohn, D.M. Crokek, *Immobilization of DNAszyme Catalytic Beacons on Pmma for Pb²⁺ Detection, Lab on a Chip* 8 (2008) 786.
- [28] Z. Wang, J.H. Lee, Y. Lu, *Label Free Colorimetric Detection of Metal Ions Using Gold Nanoparticles and DNAszyme with 3 nM Detection Limit and Tunable Dynamic Range*, Adv. Mater. 20 (2008) 3263.
- [29] H. Wei, B. Li, J. Li, S. Dong, E. Wang, *DNAszyme-Based Colorimetric Sensing of Lead (Pb²⁺) Using Unmodified Gold Nanoparticle Probes*, Nanotechnology 19 (2008) 095501/1.
- [30] A.K. Brown, J. Liu, Y. He, Y. Lu, *Biochemical Characterization of a Uranyl Ion-Specific DNAszyme*, ChemBioChem 10 (2009) 486.
- [31] S.M. Nimjee, C.P. Rusconi, B.A. Sullenger, *Aptamers: An Emerging Class of Therapeutics*, Annu. Rev. Med. 56 (2005) 555.
- [32] S.M. Shamah, J.M. Healy, S.T. Cload, *Complex Target Selex*, Acc. Chem. Res. 41 (2008) 130.
- [33] M.N. Stojanovic, P. de Prada, D.W. Landry, *Fluorescent Sensors Based on Aptamer Self-Assembly*, *J. Am. Chem. Soc.* 122 (2000) 11547.
- [34] H. Saitoh, A. Nakamura, M. Kuwahara, H. Ozaki, H. Sawai, *Modified DNA Aptamers against Sweet Agent Aspartame*, Nucl. Acids Res. Suppl. 2 (2002) 215.
- [35] D. Jellinek, L.S. Green, C. Bell, C.K. Lynott, N. Gill, C. Vargeese, G. Kirschenheuter, D.P.C. McGee, P. Abesinghe, et al., *Potent 2'-Amino-2'-Deoxypyrimidine RNA Inhibitors of Basic Fibroblast Growth Factor*, Biochemistry 34 (1995) 11363.
- [36] N.C. Pagratis, C. Bell, Y.-F. Chang, S. Jennings, T. Fitzwater, D. Jellinek, C. Dang, *Potent 2'-Amino-, and 2'-Fluoro-2'-Deoxyribonucleotide RNA Inhibitors of Keratinocyte Growth Factor*, Nat. Biotechnol. 15 (1997) 68.
- [37] J. Ruckman, L.S. Green, J. Beeson, S. Waugh, W.L. Gillette, D.D. Henninger, L. Claesson-Welsh, N. Janjic, *2'-Fluoropyrimidine RNA-Based Aptamers to the 165-Amino Acid Form of Vascular Endothelial Growth Factor (VEGF165). Inhibition of Receptor Binding and VEGF-Induced Vascular Permeability through Interactions Requiring the Exon 7-Encoded Domain*, *J. Biol. Chem.* 273 (1998) 20556.
- [38] L.S. Green, D. Jellinek, R. Jenison, A. Oestman, C.-H. Heldin, N. Janjic, *Inhibitory DNA Ligands to Platelet-Derived Growth Factor B-Chain*, Biochemistry 35 (1996) 14413.
- [39] A. Ogawa, N. Tomita, N. Kikuchi, S. Sando, Y. Aoyama, *Aptamer Selection for the Inhibition of Cell Adhesion with Fibronectin as Target*, Bioorg. Med. Chem. Lett. 14 (2004) 4001.
- [40] S.D. Mendonsa, M.T. Bowser, *In Vitro Selection of Aptamers with Affinity for Neuropeptide Y Using Capillary Electrophoresis*, *J. Am. Chem. Soc.* 127 (2005) 9382.
- [41] J. Tang, J. Xie, N. Shao, Y. Yan, *The DNA Aptamers That Specifically Recognize Ricin Toxin Are Selected by Two in Vitro Selection Methods*, Electrophoresis 27 (2006) 1303.
- [42] J. Tang, T. Yu, L. Guo, J. Xie, N. Shao, Z. He, *In Vitro Selection of DNA Aptamer against Abrin Toxin and Aptamer-Based Abrin Direct Detection*, Biosens. Bioelectron. 22 (2007) 2456.
- [43] S.H. Jeon, B. Kayhan, T. Ben-Yedidia, R. Arnon, *A DNA Aptamer Prevents Influenza Infection by Blocking the Receptor Binding Region of the Viral Hemagglutinin*, *J. Biol. Chem.* 279 (2004) 48410.
- [44] T.H. Koch, D. Smith, E. Tabacman, D.A. Zichi, *Kinetic Analysis of Site-Specific Photoaptamer-Protein Cross-Linking*, *J. Mol. Biol.* 336 (2004) 1159.
- [45] J.G. Bruno, J.L. Kiel, *In Vitro Selection of DNA Aptamers to Anthrax Spores with Electrochemiluminescence Detection*, Biosens. Bioelectron. 14 (1999) 457.
- [46] M. Blank, T. Weinschenk, M. Priemer, H. Schluesener, *Systematic Evolution of a DNA Aptamer Binding to Rat Brain Tumor Microvessels. Selective Targeting of Endothelial Regulatory Protein Pigen*, *J. Biol. Chem.* 276 (2001) 16464.
- [47] C. Wang, M. Zhang, G. Yang, D. Zhang, H. Ding, H. Wang, M. Fan, B. Shen, N. Shao, *Single-Stranded DNA Aptamers That Bind Differentiated but Not Parental Cells: Subtractive Systematic Evolution of Ligands by Exponential Enrichment*, *J. Biotechnol.* 102 (2003) 15.
- [48] E.S. Gragoudas, A.P. Adamis, E.T. Cunningham Jr, M. Feinsod, D.R. Guyer, *VEGF Inhibition Study in Ocular Neovascularization Clinical Trial Group*, Pegaptanib for Neovascular Age-Related Macular Degeneration, N. Engl. J. Med. 351 (2004) 2805.
- [49] E.W. Ng, D.T. Shima, P. Calias, E.T. Cunningham Jr, D.R. Guyer, A.P. Adamis, *Pegaptanib, a Targeted Anti-VEGF Aptamer for Ocular Vascular Disease*, Nat. Rev. Drug Discov. 5 (2006) 123.
- [50] O.C. Farokhzad, J. Cheng, B.A. Teply, I. Sherifi, S. Jon, P.W. Kantoff, J.P. Richie, R. Langer, *Targeted Nanoparticle-Aptamer Bioconjugates for Cancer Chemotherapy in Vivo*, PNAS 103 (2006) 6315.
- [51] D. Proske, M. Blank, R. Buhmann, A. Resch, *Aptamers-Basic Research, Drug Development, and Clinical Applications*, Appl. Microbiol. Biotechnol. 69 (2005) 367.
- [52] Y. Fichou, C. Ferec, *The Potential of Oligonucleotides for Therapeutic Applications*, Trends Biotechnol. 24 (2006) 563.
- [53] H. Ulrich, C.A. Trujillo, A.A. Nery, J.M. Alves, P. Majumder, R.R. Resende, A.H. Martins, *DNA and RNA Aptamers: From Tools for Basic Research Towards Therapeutic Applications*, Comb. Chem. High T. Scr. 9 (2006) 619.
- [54] S. Missailidis, A. Perkins, *Update: Aptamers as Novel Radiopharmaceuticals: Their Applications and Future Prospects in Diagnosis and Therapy*, Cancer Biother. Radiopharm. 22 (2007) 453.
- [55] S. Tombelli, M. Minunni, M. Mascini, *Aptamers-Based Assays for Diagnostics, Environmental and Food Analysis*, Biomol. Eng. 24 (2007) 191.
- [56] M. Famulok, J.S. Hartig, G. Mayer, *Functional Aptamers and Aptazymes in Biotechnology, Diagnostics, and Therapy*, Chem. Rev. 107 (2007) 3715.
- [57] S. Balamurugan, A. Obubufo, S.A. Soper, D.A. Spivak, *Surface Immobilization Methods for Aptamer Diagnostic Applications*, Anal. Bioanal. Chem. 390 (2008) 1009.
- [58] A.D. Ellington, J.W. Szostak, *In Vitro Selection of RNA Molecules That Bind Specific Ligands*, Nature 346 (1990) 818.
- [59] C. Tuerk, L. Gold, *Systematic Evolution of Ligands by Exponential Enrichment: RNA Ligands to Bacteriophage T4 DNA Polymerase*, Science 249 (1990) 505.

- [60] D. Shangguan, Y. Li, Z. Tang, Z.C. Cao, H.W. Chen, P. Mallikaratchy, K. Sefah, C.J. Yang, W. Tan, Aptamers Evolved from Live Cells as Effective Molecular Probes for Cancer Study, *PNAS* 103 (2006) 11838.
- [61] L.C. Bock, L.C. Griffin, J.A. Latham, E.H. Vermaas, J.J. Toole, Selection of Single-Stranded DNA Molecules That Bind and Inhibit Human Thrombin, *Nature* 355 (1992) 564.
- [62] J.K. Herr, J.E. Smith, C.D. Medley, D. Shangguan, W. Tan, Aptamer-Conjugated Nanoparticles for Selective Collection and Detection of Cancer Cells, *Anal. Chem.* 78 (2006) 2918.
- [63] M. Sassanfar, J.W. Szostak, An RNA Motif That Binds Atp, *Nature* 364 (1993) 550.
- [64] M. Meli, J. Vergne, J.-L. Decout, M.-C. Maurel, Adenine-Aptamer Complexes. A Bipartite RNA Site That Binds the Adenine Nucleic Base, *J. Biol. Chem.* 277 (2002) 2104.
- [65] A.D. Ellington, J.W. Szostak, Selection in Vitro of Single-Stranded DNA Molecules That Fold into Specific Ligand-Binding Structures, *Nature* 355 (1992) 850.
- [66] M. Famulok, J.W. Szostak, Stereospecific Recognition of Tryptophan Agarose by In Vitro Selected RNA, *J. Am. Chem. Soc.* 114 (1992) 3990.
- [67] J. Ciesiolka, J. Gorski, M. Yarus, Selection of an RNA Domain That Binds Zn^{2+} , *RNA* 1 (1995) 538.
- [68] J. Ciesiolka, M. Yarus, Small RNA-Divalent Domains, *RNA* 2 (1996) 785.
- [69] J.H. Lee, D.P. Wernette, M.V. Yigit, J. Liu, Z. Wang, Y. Lu, Site-Specific Control of Distances between Gold Nanoparticles Using Phosphorothioate Anchors on DNA and a Short Bifunctional Molecular Fastener, *Angew. Chem. Int. Ed.* 46 (2007) 9006.
- [70] E. Winfree, F. Liu, L.A. Wenzler, N.C. Seeman, Design and Self-Assembly of Two-Dimensional DNA Crystals, *Nature* 394 (1998) 539.
- [71] R.P. Goodman, I.A. Schaap, C.F. Tardin, C.M. Erben, R.M. Berry, C.F. Schmidt, A.J. Turberfield, Rapid Chiral Assembly of Rigid DNA Building Blocks for Molecular Nanofabrication, *Science* 310 (2005) 1661.
- [72] P.W.K. Rothmund, Folding DNA to Create Nanoscale Shapes and Patterns, *Nature* 440 (2006) 297.
- [73] Y. He, T. Ye, M. Su, C. Zhang, A.E. Ribbe, W. Jiang, C. Mao, Hierarchical Self-Assembly of DNA into Symmetric Supramolecular Polyhedra, *Nature* 452 (2008) 198.
- [74] N.C. Seeman, DNA in a Material World, *Nature* 421 (2003) 427.
- [75] N.C. Seeman, The Design and Engineering of Nucleic Acid Nanoscale Assemblies, *Curr. Opin. Struct. Biol.* 6 (1996) 519.
- [76] Y. Ke, S. Lindsay, Y. Chang, Y. Liu, H. Yan, Self-Assembled Water-Soluble Nucleic Acid Probe Tiles for Label-Free RNA Hybridization Assays, *Science* 319 (2008) 180.
- [77] S. Rinker, Y. Ke, Y. Liu, R. Chhabra, H. Yan, Self-Assembled DNA Nanostructures for Distance-Dependent Multivalent Ligand-Protein Binding, *Nat. Nanotechnol* 3 (2008) 418.
- [78] J. Clark, S.S. Smith, Application of Nanoscale Bioassemblies to Clinical Laboratory Diagnostics, *Adv. Clin. Chem.* 41 (2006) 23.
- [79] I. Willner, M. Zayats, Electronic Aptamer-Based Sensors, *Angew. Chem. Int. Ed.* 46 (2007) 6408.
- [80] R. Langer, Drug Delivery and Targeting, *Nature* 392 (1998) 5.
- [81] C. Wilson, J.W. Szostak, Isolation of a Fluorophore-Specific DNA Aptamer with Weak Redox Activity, *Chem. Biol.* 5 (1998) 609.
- [82] M. Liss, B. Petersen, H. Wolf, E. Prohaska, An Aptamer-Based Quartz Crystal Protein Biosensor, *Anal. Chem.* 74 (2002) 4488.
- [83] L. Gold, B. Polisky, O. Uhlenbeck, M. Yarus, Diversity of Oligonucleotide Functions, *Annu. Rev. Biochem.* 64 (1995) 763.
- [84] P.E. Burmeister, S.D. Lewis, R.F. Silva, J.R. Preiss, L.R. Horwitz, P.S. Pendergrast, T.G. McCauley, J.C. Kurz, D.M. Epstein, C. Wilson, A.D. Keefe, Direct in Vitro Selection of a 2'-O-Methyl Aptamer to VEGF, *Chem. Biol.* 12 (2005) 25.
- [85] J.-U. Park, J.H. Lee, U. Paik, Y. Lu, J.A. Rogers, Nanoscale Patterns of Oligonucleotides Formed by Electrohydrodynamic Jet Printing with Applications in Biosensing and Nanomaterials Assembly, *Nano Lett.* 8 (2008) 4210.
- [86] C.B. Murray, D.J. Norris, M.G. Bawendi, Synthesis and Characterization of Nearly Monodisperse CdE (E = Sulfur, Selenium, Tellurium) Semiconductor Nanocrystallites, *J. Am. Chem. Soc.* 115 (1993) 8706.
- [87] A.P. Alivisatos, Semiconductor Clusters, Nanocrystals, and Quantum Dots, *Science* 271 (1996) 933.
- [88] X. Peng, L. Manna, W. Yang, J. Wickham, E. Scher, A. Kadavanich, A.P. Alivisatos, Shape Control of CdSe Nanocrystals, *Nature* 404 (2000) 59.
- [89] G.P. Mitchell, C.A. Mirkin, R.L. Letsinger, Programmed Assembly of DNA Functionalized Quantum Dots, *J. Am. Chem. Soc.* 121 (1999) 8122.
- [90] R.Q. Liang, W. Li, Y. Li, C.Y. Tan, J.X. Li, Y.X. Jin, K.C. Ruan, An Oligonucleotide Microarray for MicroRNA Expression Analysis Based on Labeling RNA with Quantum Dot and Nanogold Probe, *Nucleic Acids Res.* 33 (2005) e17.
- [91] C.A. Lin, R.A. Sperling, J.K. Li, T.Y. Yang, P.Y. Li, M. Zanella, W.H. Chang, W.J. Parak, Design of an Amphiphilic Polymer for Nanoparticle Coating and Functionalization, *Small* 4 (2008) 334.
- [92] C.Y. Zhang, H.C. Yeh, M.T. Kuroki, T.H. Wang, Single-Quantum-Dot-Based DNA Nanosensor, *Nat. Mater.* 4 (2005) 826.
- [93] R. Gill, I. Willner, I. Shweky, U. Banin, Fluorescence Resonance Energy Transfer in CdSe/ZnS-DNA Conjugates: Probing Hybridization and DNA Cleavage, *J. Phys. Chem. B* 109 (2005) 23715.
- [94] M. Levy, S.F. Cater, A.D. Ellington, Quantum-Dot Aptamer Beacons for the Detection of Proteins, *ChemBioChem* 6 (2005) 2163.
- [95] J.H. Choi, K.H. Chen, M.S. Strano, Aptamer-Capped Nanocrystal Quantum Dots: A New Method for Label-Free Protein Detection, *J. Am. Chem. Soc.* 128 (2006) 15584.
- [96] J. Liu, J.H. Lee, Y. Lu, Quantum Dot Encoding of Aptamer-Linked Nanostructures for One Pot Simultaneous Detection of Multiple Analytes, *Anal. Chem.* 79 (2007) 4120.
- [97] P. Cherukuri, S.M. Bachilo, S.H. Litovsky, R.B. Weisman, Near-Infrared Fluorescence Microscopy of Single-Walled Carbon Nanotubes in Phagocytic Cells, *J. Am. Chem. Soc.* 126 (2004) 15638.
- [98] Z. Zhu, Z. Tang, J.A. Phillips, R. Yang, H. Wang, W. Tan, Regulation of Singlet Oxygen Generation Using Single-Walled Carbon Nanotubes, *J. Am. Chem. Soc.* 130 (2008) 10856.
- [99] R. Yang, Z. Tang, J. Yan, H. Kang, Y. Kim, Z. Zhu, W. Tan, Noncovalent Assembly of Carbon Nanotubes and Single-Stranded DNA: An Effective Sensing Platform for Probing Biomolecular Interactions, *Anal. Chem.* 80 (2008) 7408.
- [100] C.B. Swearingen, D.P. Wernette, D.M. Cropek, Y. Lu, J.V. Sweedler, P.W. Bohn, Immobilization of a Catalytic DNA Molecular Beacon on Au for Pb(II) Detection, *Anal. Chem.* 77 (2005) 442.
- [101] D.P. Wernette, C.B. Swearingen, D.M. Cropek, Y. Lu, J.V. Sweedler, P.W. Bohn, Incorporation of a DNAzyme into Au-Coated Nanocapillary Array Membranes with an Internal Standard for Pb(II) Sensing, *Analyst* 131 (2006) 41.
- [102] D.P. Wernette, C. Mead, P.W. Bohn, Y. Lu, Surface Immobilization of Catalytic Beacons Based on Ratiometric Fluorescent DNAzyme Sensors: A Systematic Study, *Langmuir* 23 (2007) 9513.
- [103] T.C. Kuo, D.M. Cannon, M.A. Shannon, P.W. Bohn, J.V. Sweedler, Hybrid Three-Dimensional Nanofluidic/Microfluidic Devices Using Molecular Gates, *Sens. Actuators, A* 102 (2003) 223.
- [104] T.C. Kuo, D.M. Cannon, Y.N. Chen, J.J. Tulock, M.A. Shannon, J.V. Sweedler, P.W. Bohn, Gateable Nanofluidic Interconnects for Multilayered Microfluidic Separation Systems, *Anal. Chem.* 75 (2003) 1861.
- [105] L.-H. Chang, J.J. Tulock, J. Liu, W.-S. Kim, D.M. Cannon Jr, Y. Lu, P.W. Bohn, J.V. Sweedler, D.M. Cropek, Miniaturized Lead Sensor Based on Lead-Specific DNAzyme in a Nanocapillary Interconnected Microfluidic Device, *Environ. Sci. Technol.* 39 (2005) 3756.
- [106] K.A. Shaikh, K.S. Ryu, E.D. Goluch, J.-M. Nam, J. Liu, C.S. Thaxton, T.N. Chiesl, A.E. Barron, Y. Lu, C.A. Mirkin, C. Liu, A Modular Microfluidic Architecture for Integrated Biochemical Analysis, *PNAS* 102 (2005) 9745.
- [107] M.N. Stojanovic, D.W. Landry, Aptamer-Based Colorimetric Probe for Cocaine, *J. Am. Chem. Soc.* 124 (2002) 9678.
- [108] V. Pavlov, Y. Xiao, B. Shlyahovskiy, I. Willner, Aptamer-Functionalized Au Nanoparticles for the Amplified Optical Detection of Thrombin, *J. Am. Chem. Soc.* 126 (2004) 11768.
- [109] H.-A. Ho, M. Leclerc, Optical Sensors Based on Hybrid Aptamer/Conjugated Polymer Complexes, *J. Am. Chem. Soc.* 126 (2004) 1384.
- [110] C.-C. Huang, Y.-F. Huang, Z. Cao, W. Tan, H.-T. Chang, Aptamer-Modified Gold Nanoparticles for Colorimetric Determination of Platelet-Derived Growth Factors and Their Receptors, *Anal. Chem.* 77 (2005) 5735.
- [111] J. Liu, Y. Lu, Fast Colorimetric Sensing of Adenosine and Cocaine Based on a General Sensor Design Involving Aptamers and Nanoparticles, *Angew. Chem. Int. Ed.* 45 (2006) 90.
- [112] J. Chen, F. Saeki, B.J. Wiley, H. Cang, M.J. Cobb, Z.-Y. Li, L. Au, H. Zhang, M.B. Kimmey, X. Li, Y. Xia, Gold Nanocages: Bioconjugation and Their Potential Use as Optical Imaging Contrast Agents, *Nano Lett.* 5 (2005) 473.
- [113] C.A. Mirkin, R.L. Letsinger, R.C. Mucic, J.J. Storhoff, A DNA-Based Method for Rationally Assembling Nanoparticles into Macroscopic Materials, *Nature* 382 (1996) 607.
- [114] H. Li, L.J. Rothberg, Label-Free Colorimetric Detection of Specific Sequences in Genomic DNA Amplified by the Polymerase Chain Reaction, *J. Am. Chem. Soc.* 126 (2004) 10958.
- [115] J.H. Lee, Z. Wang, J. Liu, Y. Lu, Highly Sensitive and Selective Colorimetric Sensors for Uranyl (UO_2^{2+}): Development and Comparison of Labeled and Label-Free DNAzyme-Gold Nanoparticle Systems, *J. Am. Chem. Soc.* 130 (2008) 14217.
- [116] Y. Wang, D. Li, W. Ren, Z. Liu, S. Dong, E. Wang, Ultrasensitive Colorimetric Detection of Protein by Aptamer-Au Nanoparticles Conjugates Based on a Dot-Blot Assay, *Chem. Commun.* 22 (2008) 2520.
- [117] J. Liu, Y. Lu, Smart Nanomaterials Responsive to Multiple Chemical Stimuli with Controllable Cooperativity, *Adv. Mater.* 18 (2006) 1667.
- [118] J. Wang, L. Wang, X. Liu, Z. Liang, S. Song, W. Li, G. Li, C. Fan, A Gold Nanoparticle-Based Aptamer Target Binding Readout for ATP Assay, *Adv. Mater. Des.* 19 (2007) 3943.
- [119] H. Wei, B. Li, J. Li, E. Wang, S. Dong, Simple and Sensitive Aptamer-Based Colorimetric Sensing of Protein Using Unmodified Gold Nanoparticle Probes, *Chem. Commun.* 36 (2007) 3735.
- [120] W. Zhao, W. Chiuman, M.A. Brook, Y. Li, Simple and Rapid Colorimetric Biosensors Based on DNA Aptamer and Noncrosslinking Gold Nanoparticle Aggregation, *ChemBioChem* 8 (2007) 727.
- [121] W. Zhao, W. Chiuman, J.C.F. Lam, S.A. McManus, W. Chen, Y. Cui, R. Pelton, M.A. Brook, Y. Li, DNA Aptamer Folding on Gold Nanoparticles: From Colloid Chemistry to Biosensors, *J. Am. Chem. Soc.* 130 (2008) 3610.
- [122] C.P. Chan, Y.C. Cheung, R. Renneberg, M. Seydack, New Trends in Immunoassays, *Adv. Biochem. Eng. Biotechnol.* 109 (2008) 123.
- [123] K. Glynou, P.C. Ioannou, T.K. Christopoulos, V. Syriopoulou, Oligonucleotide-Functionalized Gold Nanoparticles as Probes in a Dry-Reagent Strip Biosensor for DNA Analysis by Hybridization, *Anal. Chem.* 75 (2003) 4155.
- [124] J. Liu, D. Mazumdar, Y. Lu, A Simple and Sensitive "Dipstick" Test in Serum Based on Lateral Flow Separation of Aptamer-Linked Nanostructures, *Angew. Chem. Int. Ed.* 45 (2006) 7955.
- [125] S. Su, R. Nutui, C.D.M. Filipe, Y. Li, R. Pelton, Adsorption and Covalent Coupling of ATP-Binding DNA Aptamers onto Cellulose, *Langmuir* 23 (2007) 1300.
- [126] S. Nie, S.R. Emory, Probing Single Molecules and Single Nanoparticles by Surface-Enhanced Raman Scattering, *Science* 275 (1997) 1102.

- [127] M. Fleischmann, P.J. Hendra, A.J. McQuillan, Raman Spectra of Pyridine Adsorbed at a Silver Electrode, *Chem. Phys. Lett.* 26 (1974) 163.
- [128] K.A. Willlets, R.P. Van Duyne, Localized Surface Plasmon Resonance Spectroscopy and Sensing, *Annu. Rev. Phys. Chem.* 58 (2007) 267.
- [129] K. Kneipp, Y. Wang, H. Kneipp, L.T. Perelman, I. Itzkan, R.R. Dasari, M.S. Feld, Single molecule detection using surface-enhanced raman scattering, *Phys. Rev. Lett.* 78 (1997) 1667.
- [130] Y.C. Cao, R. Jin, C.A. Mirkin, Nanoparticles with raman spectroscopic fingerprints for DNA and RNA detection, *Science* 297 (2002) 1536.
- [131] A. Barhoumi, D. Zhang, F. Tam, N.J. Halas, Surface-enhanced raman spectroscopy of DNA, *J. Am. Chem. Soc.* 130 (2008) 5523.
- [132] Y. Wang, H. Wei, B. Li, W. Ren, S. Guo, S. Dong, E. Wang, Sers Opens a New Way in Aptasensor for Protein Recognition with High Sensitivity and Selectivity, *Chem. Commun.* 48 (2007) 5220.
- [133] D.M. Tasset, M.F. Kubik, W. Steiner, Oligonucleotide Inhibitors of Human Thrombin That Bind Distinct Epitopes, *J. Mol. Biol.* 272 (1997) 688.
- [134] J.W. Chen, X.P. Liu, K.J. Feng, Y. Liang, J.H. Jiang, G.L. Shen, R.Q. Yu, Detection of Adenosine Using Surface-Enhanced Raman Scattering Based on Structure-Switching Signaling Aptamer, *Biosens. Bioelectron.* 24 (2008) 66.
- [135] X. Qian, X.H. Peng, D.O. Ansari, Q. Yin-Goen, G.Z. Chen, D.M. Shin, L. Yang, A.N. Young, M.D. Wang, S. Nie, In Vivo Tumor Targeting and Spectroscopic Detection with Surface-Enhanced Raman Nanoparticle Tags, *Nat. Biotechnol.* 26 (2008) 83.
- [136] S. Keren, C. Zavaleta, Z. Cheng, A. de la Zerda, O. Gheysens, S.S. Gambhir, Noninvasive Molecular Imaging of Small Living Subjects Using Raman Spectroscopy, *PNAS* 105 (2008) 5844.
- [137] L. Josephson, J.M. Perez, R. Weissleder, Magnetic Nanosensors for the Detection of Oligonucleotide Sequences, *Angew. Chem. Int. Ed.* 40 (2001) 3204.
- [138] J.M. Perez, L. Josephson, T. O'Loughlin, D. Hoegemann, R. Weissleder, Magnetic Relaxation Switches Capable of Sensing Molecular Interactions, *Nat. Biotechnol.* 20 (2002) 816.
- [139] M. Zhao, L. Josephson, Y. Tang, R. Weissleder, Magnetic Sensors for Protease Assays, *Angew. Chem. Int. Ed.* 42 (2003) 1375.
- [140] A. Tsourkas, O. Hofstetter, H. Hofstetter, R. Weissleder, L. Josephson, Magnetic Relaxation Switch Immunosenors Detect Enantiomeric Impurities, *Angew. Chem. Int. Ed.* 43 (2004) 2395.
- [141] J.M. Perez, L. Josephson, R. Weissleder, Use of Magnetic Nanoparticles as Nanosensors to Probe for Molecular Interactions, *ChemBioChem* 5 (2004) 261.
- [142] M.V. Yigit, D. Mazumdar, H.K. Kim, J.H. Lee, B. Odintsov, Y. Lu, Smart "Turn-On" Magnetic Resonance Contrast Agents Based on Aptamer-Functionalized Superparamagnetic Iron Oxide Nanoparticles, *ChemBioChem* 8 (2007) 1675.
- [143] M.V. Yigit, D. Mazumdar, Y. Lu, Mri Detection of Thrombin with Aptamer Functionalized Superparamagnetic Iron Oxide Nanoparticles, *Bioconjug. Chem.* 19 (2008) 412.
- [144] E. Katz, I. Willner, Biomolecule-Functionalized Carbon Nanotubes: Applications in Nanobioelectronics, *ChemPhysChem* 5 (2004) 1084.
- [145] F. Patolsky, E. Katz, I. Willner, Amplified DNA Detection by Electrogenenerated Biochemiluminescence and by the Catalyzed Precipitation of an Insoluble Product on Electrodes in the Presence of the Doxorubicin Intercalator, *Angew. Chem. Int. Ed.* 41 (2002) 3398.
- [146] F. Patolsky, A. Lichtenstein, I. Willner, Highly Sensitive Amplified Electronic Detection of DNA by Biocatalyzed Precipitation of an Insoluble Product onto Electrodes, *Chem. Eur. J.* 9 (2003) 1137.
- [147] F. Patolsky, A. Lichtenstein, I. Willner, Detection of Single-Base DNA Mutations by Enzyme-Amplified Electronic Transduction, *Nat. Biotechnol.* 19 (2001) 253.
- [148] R. Polsky, R. Gill, L. Kaganovsky, I. Willner, Nucleic Acid-Functionalized Pt Nanoparticles: Catalytic Labels for the Amplified Electrochemical Detection of Biomolecules, *Anal. Chem.* 78 (2006) 2268.
- [149] M. Mir, M. Vreeke, I. Katakis, Different Strategies to Develop an Electrochemical Thrombin Aptasensor, *Electrochem. Commun.* 8 (2006) 505.
- [150] K. Ikebukuro, C. Kiyohara, K. Sode, Novel Electrochemical Sensor System for Protein Using the Aptamers in Sandwich Manner, *Biosens. Bioelectron.* 20 (2005) 2168.
- [151] B.R. Baker, R.Y. Lai, M.S. Wood, E.H. Doctor, A.J. Heeger, K.W. Plaxco, An Electronic, Aptamer-Based Small-Molecule Sensor for the Rapid, Label-Free Detection of Cocaine in Adulterated Samples and Biological Fluids, *J. Am. Chem. Soc.* 128 (2006) 3138.
- [152] Z.-S. Wu, M.-M. Guo, S.-B. Zhang, C.-R. Chen, J.-H. Jiang, G.-L. Shen, R.-Q. Yu, Reusable Electrochemical Sensing Platform for Highly Sensitive Detection of Small Molecules Based on Structure-Switching Signaling Aptamers, *Anal. Chem.* 79 (2007) 2933.
- [153] X. Zuo, S. Song, J. Zhang, D. Pan, L. Wang, C. Fan, A Target-Responsive Electrochemical Aptamer Switch (TREAS) for Reagentless Detection of Nanomolar Atp, *J. Am. Chem. Soc.* 129 (2007) 1042.
- [154] Y. Xiao, A.A. Lubin, A.J. Heeger, K.W. Plaxco, Label-Free Electronic Detection of Thrombin in Blood Serum by Using an Aptamer-Based Sensor, *Angew. Chem. Int. Ed.* 44 (2005) 5456.
- [155] A.-E. Radi, J.L. Acero Sanchez, E. Baldrich, C.K. O'Sullivan, Reagentless, Reusable, Ultrasensitive Electrochemical Molecular Beacon Aptasensor, *J. Am. Chem. Soc.* 128 (2006) 117.
- [156] Y. Xiao, B.D. Piorek, K.W. Plaxco, A.J. Heeger, A Reagentless Signal-on-Architecture for Electronic, Aptamer-Based Sensors Via Target-Induced Strand Displacement, *J. Am. Chem. Soc.* 127 (2005) 17990.
- [157] Y. Du, B. Li, H. Wei, Y. Wang, E. Wang, Multifunctional Label-Free Electrochemical Biosensor Based on an Integrated Aptamer, *Anal. Chem.* 80 (2008) 5110.
- [158] J.A. Hansen, J. Wang, A.-N. Kawde, Y. Xiang, K.V. Gothelf, G. Collins, Quantum-Dot/Aptamer-Based Ultrasensitive Multi-Analyte Electrochemical Biosensor, *J. Am. Chem. Soc.* 128 (2006) 2228.
- [159] M. Zayats, Y. Huang, R. Gill, C.-a. Ma, I. Willner, Label-Free and Reagentless Aptamer-Based Sensors for Small Molecules, *J. Am. Chem. Soc.* 128 (2006) 13666.
- [160] S. Iijima, Helical Microtubules of Graphitic Carbon, *Nature* 354 (1991) 56.
- [161] P.M. Ajayan, Nanotubes from Carbon, *Chem. Rev.* 99 (1999) 1787.
- [162] H. Dai, Carbon Nanotubes: Synthesis, Integration, and Properties, *Acc. Chem. Res.* 35 (2002) 1035.
- [163] N.W. Kam, Z. Liu, H. Dai, Carbon Nanotubes as Intracellular Transporters for Proteins and DNA: An Investigation of the Uptake Mechanism and Pathway, *Angew. Chem. Int. Ed.* 45 (2006) 577.
- [164] C.V. Nguyen, L. Delzeit, A.M. Cassell, J. Li, J. Han, M. Meyyappan, Preparation of Nucleic Acid Functionalized Carbon Nanotube Arrays, *Nano Lett.* 2 (2002) 1079.
- [165] M. Hazani, R. Naaman, F. Hennrich, M.M. Kappes, Confocal Fluorescence Imaging of DNA-Functionalized Carbon Nanotubes, *Nano Lett.* 3 (2003) 153.
- [166] C. Dwyer, M. Guthold, M. Falvo, S. Washburn, R. Superfine, D. Eri, DNA-Functionalized Single-Walled Carbon Nanotubes, *Nanotechnology* 13 (2002) 601.
- [167] S.E. Baker, W. Cai, T.L. Lasseter, K.P. Weidkamp, R.J. Hamers, Covalently Bonded Adducts of Deoxyribonucleic Acid (DNA) Oligonucleotides with Single-Wall Carbon Nanotubes: Synthesis and Hybridization, *Nano Lett.* 2 (2002) 1413.
- [168] R.J. Chen, Y. Zhang, D. Wang, H. Dai, Noncovalent Sidewall Functionalization of Single-Walled Carbon Nanotubes for Protein Immobilization, *J. Am. Chem. Soc.* 123 (2001) 3838.
- [169] R.J. Chen, S. Bangsaruntip, K.A. Drouvalakis, N.W.S. Kam, M. Shim, Y. Li, W. Kim, P.J. Utz, H. Dai, Noncovalent Functionalization of Carbon Nanotubes for Highly Specific Electronic Biosensors, *PNAS* 100 (2003) 4984.
- [170] H.-M. So, K. Won, Y.H. Kim, B.-K. Kim, B.H. Ryu, P.S. Na, H. Kim, J.-O. Lee, Single-Walled Carbon Nanotube Biosensors Using Aptamers as Molecular Recognition Elements, *J. Am. Chem. Soc.* 127 (2005) 11906.
- [171] K. Maehashi, T. Katsura, K. Kerman, Y. Takamura, K. Matsumoto, E. Tamiya, Label-Free Protein Biosensor Based on Aptamer-Modified Carbon Nanotube Field-Effect Transistors, *Anal. Chem.* 79 (2007) 782.
- [172] H.M. So, D.W. Park, E.K. Jeon, Y.H. Kim, B.S. Kim, C.K. Lee, S.Y. Choi, S.C. Kim, H. Chang, J.O. Lee, Detection and Titer Estimation of Escherichia Coli Using Aptamer-Functionalized Single-Walled Carbon-Nanotube Field-Effect Transistors, *Small* 4 (2008) 197.
- [173] V. Bagalkot, L. Zhang, E. Levy-Nissenbaum, S. Jon, P.W. Kantoff, R. Langer, O.C. Farokhzad, Quantum Dot-Aptamer Conjugates for Synchronous Cancer Imaging, Therapy, and Sensing of Drug Delivery Based on Bi-Fluorescence Resonance Energy Transfer, *Nano Lett.* 7 (2007) 3065.
- [174] V. Bagalkot, O.C. Farokhzad, R. Langer, S. Jon, An Aptamer-Doxorubicin Physical Conjugate as a Novel Targeted Drug-Delivery Platform, *Angew. Chem. Int. Ed.* 45 (2006) 8149.
- [175] A.Z. Wang, V. Bagalkot, C.C. Vasiliou, F. Gu, F. Alexis, L. Zhang, M. Shaikh, K. Yuet, M.J. Cima, R. Langer, P.W. Kantoff, N.H. Bander, S. Jon, O.C. Farokhzad, Superparamagnetic Iron Oxide Nanoparticle-Aptamer Bioconjugates for Combined Prostate Cancer Imaging and Therapy, *ChemMedChem* 3 (2008) 1311.
- [176] S. Dhar, F.X. Gu, R. Langer, O.C. Farokhzad, S.J. Lippard, Targeted Delivery of Cisplatin to Prostate Cancer Cells by Aptamer Functionalized Pt(IV) Prodrug-PLGA-PEG Nanoparticles, *PNAS* 105 (2008) 17356.
- [177] Y.-F. Huang, H.-T. Chang, W. Tan, Cancer Cell Targeting Using Multiple Aptamers Conjugated on Nanorods, *Anal. Chem.* 80 (2008) 567.
- [178] Y. Wu, J.A. Phillips, H. Liu, R. Yang, W. Tan, Carbon Nanotubes Protect DNA Strands During Cellular Delivery, *ACS Nano* 2 (2008) 2023.
- [179] G.J. Tong, S.C. Hsiao, Z.M. Carrico, M.B. Francis, Viral Capsid DNA Aptamer Conjugates as Multivalent Cell-Targeting Vehicles, *J. Am. Chem. Soc.* 131 (2009) 11174.
- [180] Z. Cao, R. Tong, A. Mishra, W. Xu, G.C.L. Wong, J. Cheng, Y. Lu, Reversible Cell-Specific Drug Delivery with-Functionalized Liposomes, *Angew. Chem. Int. Ed.* 48 (2009) 6494.